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Construction of polycyclic fused pyrrolidines with three contiguous stereocentres via Michael addition of vinyl malononitriles with nitrostyrenes using L-Proline derived thiourea

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I. General Remarks:

All reactions were carried out in an oven dried flask. Solvents used for reactions and column chromatography were commercial grade and distilled prior to use. Toluene and THF were dried over sodium/benzophenone, CH₂Cl₂ and CHCl₃ over CaH₂. Solvents for HPLC bought as analytical grade and used without further purification. TLC was performed on precoated Merck silica gel aluminium plates with 60_F254 indicator, visualised by irradiation with UV light. Column chromatography was performed using silica gel Merck 60-100 mesh. ¹H-NMR and ¹³C-NMR were recorded on a Bruker AV 500 MHz using CD₃OD-d₄ and CDCl₃ as solvent and multiplicity indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (doublet of doublet), dt (doublet of triplet) bs (broad singlet). Coupling constants J were reported in Hertz. High resolution mass spectra were obtained by ESI using Thermo scientific Orbitrap Elite mass spectrometer. IR spectra were recorded on a Perkin Elmer FT/IR-420 spectrometer and are reported in terms of frequency of absorption (cm⁻¹). The enantiomeric excess is obtained by HPLC analysis using a chiral stationary phase column (CHIRALPAK ADH, CHIRALCELL OD-H and Phenomenex Amylose-2. All the physical and spectroscopic data of 3a - d, 3aa - ah, 3ba, 3bh and 7a were in complete agreement with the reported literature.

II. General procedure for asymmetric vinylogous Michael addition of vinyl malononitriles to nitrostyrene

To a stirred solution of 4 (0.92 mg, 0.002mmol, 5 mol %) and vinyl malanonitriles 1 (0.1 mmol) in Toluene (1 mL), nitrostyrene 2 (0.12 mmol) was added. The solution was stirred at ambient temperature for mentioned days. After the reaction was completed (monitored by TLC), the resulting mixture was concentrated under reduced pressure and the residue was purified through column chromatography on silica gel to give the product 3.

$2 \hbox{-} ((S) \hbox{-} 2 \hbox{-} ((R) \hbox{-} 2 \hbox{-} nitro \hbox{-} 1 \hbox{-} phenylethyl) \hbox{-} 3,4 \hbox{-} dihydronaphthalen} \hbox{-} 1 (2H) \hbox{-} ylidene) malononitrile \\ 3a \hbox{:}$

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 28.5 mg (0.083mmol) with 83% yield; 91% ee determined by HPLC on AS column, 30% 2-propanol/hexane, 1.0ml/min, UV 254nm, t_{minor} =13.3 min, t_{major} =20.4 min.

2-((S)-3-((R)-2-nitro-1-phenylethyl)chroman-4 ylidene)malononitrile 3b:

General experimental procedure I was followed to prepare the Michael addition product **3b**. The desired product was obtained as white solid 27.9 mg (0.081mmol) with 81% yield; 95% ee was determined by HPLC on AS column, 30% 2-propanol/hexane, 1.0ml/min, UV 254nm, $t_{minor} = 15.14 \text{ min}$, $t_{major} = 17.3 \text{mmin}$; $[\alpha]_D^{25} - 172.3^{\circ}$ (c 0.15, CHCl₃).

2-((S)-3-((R)-2-nitro-1-phenylethyl)thiochroman-4-ylidene)malononitrile 3c:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 28.9 mg (0.080mmol) with 80% yield; 90% ee was determined by HPLC on AS column, 35% 2-propanol/hexane, 1.0ml/min, Uv 254nm, $t_{minor} = 15.78 \text{ min}$, $t_{major} = 31.04 \text{min}$.

2-((S)-2-((R)-2-nitro-1-phenylethyl)cyclohexylidene)malononitrile 3d:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 16.5 mg (0.056mmol) with 56% yield; 28% ee was determined by HPLC on AS column, 30% 2-propanol/hexane, 1.0ml/min, Uv 254nm, $t_{major} = 7.3$ min, $t_{major} = 9.3$ min.

2-((S)-2-((R)-1-(2-fluorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile 3aa:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 28.9 mg (0.080mmol) with 80% yield; 85% ee was determined by HPLC on AS column, 35% 2-propanol/hexane, 1.0ml/min, Uv 254nm, $t_{minor} = 19.6 \text{ min}$, $t_{major} = 23.3 \text{ min}$.

2-((S)-2-((R)-1-(2-chlorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile 3ab:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 29.4 mg (0.078 mmol) with 78 % yield; 90% ee was determined by HPLC on AS column, 35% 2-propanol/hexane, 1.0ml/min, Uv 254nm, $t_{minor} = 8.1$ min, $t_{major} = 10.1$ min.

2-((S)-2-((R)-1-(2-bromophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile 3ac:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 35 mg (0.083mmol) with 93% yield; 86% ee was determined by HPLC on AS column, 35% 2-propanol/hexane, 1.0ml/min, Uv 254nm, $t_{minor} = 19.0 \text{ min}$, $t_{major} = 19.9 \text{ min}$.

2-((S)-2-((R)-1-(3-chlorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile 3ad:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 30.5 mg (0.081mmol) with 81% yield; 88% ee was determined by HPLC on AS column, 35% 2-propanol/hexane, 1.0ml/min, Uv 254nm, $t_{minor} = 10.3 \text{ min}$, $t_{major} = 15.8 \text{ min}$.

2-((S)-2-((R)-1-(3-methoxyphenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-vlidene)malononitrile 3ae:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 28.0mg (0.075mmol) with 75% yield; 84% ee was determined by HPLC on AS column, 35% 2-propanol/hexane, 1.0ml/min, Uv 254nm, t_{minor} = $29.5 \text{ min}, t_{\text{major}} = 27.11 \text{ min}.$

2-((S)-2-((R)-1-(4-chlorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile 3af:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 27.1 mg (0.072mmol) with 72% yield; 88% ee was determined by HPLC on AS column, 30% 2-propanol/hexane, 1.0ml/min, UV 254nm, t_{minor}=13.5 min, $t_{\text{major}} = 18.6 \text{min}$.

2-((S)-2-((R)-1-(4-bromophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile 3ag:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 30.4 (0.072mmol) with 72% yield; 92% ee was determined by HPLC on AS column, 30% 2-propanol/hexane, 1.0ml/min, UV 254nm, t_{minor}= $14.0 \text{ min}, t_{\text{major}} = 19.4 \text{ min}.$

2-((S)-2-((R)-1-(4-fluorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile 3ah:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 29.2 mg (0.081mmol) with 81% yield; 87% ee was determined by HPLC on AS column, 30% 2-propanol/hexane, 1.0ml/min, UV 254nm, t_{minor}=16.8 min, $t_{\text{major}} = 25.4 \text{ min.}$

2-((S)-3-((R)-1-(4-methoxyphenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile 3bh:

General experimental procedure I was followed to prepare the Michael addition product 3a. The desired product was obtained as white solid 28.5 mg (0.076mmol) with 76% yield. 91% ee was determined by HPLC on AS column, 80% 2-propanol/hexane, 1.0ml/min, UV 254nm, t_{minor}=10.2 min, t_{major}=13.2 min.

2-((S)-3-((R)-1-(2-fluorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile

General experimental procedure II was followed to prepare the Michael/hemiketalization

product **3bb.** The desired product was obtained as white solid with 75% yield, mp: 183-185°C; ¹H NMR (500MHz, CDCl₃) δ (ppm) 8.27 (dd, J = 1.5, 8.0 Hz, 1H), 7.63 (td, J = 1.5, 6.0 Hz, 1H), 7.44-7.38 (m, 1H), 7.29-7.22 (m, 2H), 7.19-7.19 (m, 3H), 7.16 (dd, J = 1.5, 10Hz, 1H), 5.02(t, J = 7.5 Hz, 1H), 4.57 (dd, J = 10.5, 18.5 Hz, 1H), 4.16 (dd, J = 3.5, 18.5 Hz)12.5 Hz, 2H), 3.58(d, J = 8.5 Hz); ¹³C NMR (125 MHz, CDCl₃) δ (ppm) 165.0, 155.9, 137.6, 131.1, 127.9, 125.3, 122.4, 118.4, 116.8, 116.6, 114.8, 112.8, 79.4, 76.3, 66.8, 59.5; HRMS (ESI) calcd for $C_{20}H_{14}FN_3O_3$ [M+H]⁺: 386.0911, found: 386.0922; IR (KBr): v 3457, 2928, 2230, 1560, 1542, 1487, 1447, 1378, 1082, 1014, 832, 760 cm⁻¹; 86% ee was determined by HPLC on ADH column, 10/90% 2-propanol/hexane, 1.0mL/min, UV 254nm, t_{minor} = 19.1 min, t_{major} = 21.8 min.

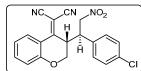
2-((S)-3-((R)-1-(3-chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile:

General experimental procedure **II** was followed to prepare the Michael addition product **3bc.** The desired product was obtained as white solid with 82% yield, mp: 187-189°C; ¹H NMR (500MHz, CDCl₃) δ = 8.28 (dd, J = 1.6, 8.2 Hz, 1 H), 7.65 (ddd, J = 1.4, 7.2, 8.4 Hz, 1 H), 7.44 - 7.38 (m, 2 H), 7.36 - 7.31 (m, 1 H), 7.27 - 7.23 (m, 1 H), 7.18 (dt, J =

1.3, 7.7 Hz, 1 H), 7.10 (dd, J = 1.1, 8.4 Hz, 1 H), 4.85 (dd, J = 10.4, 13.2 Hz, 1 H), 4.52 (dd, J = 5.2, 13.1 Hz, 1 H), 4.18 (dd, J = 2.4, 12.5 Hz, 1 H), 4.09 (dd, J = 1.7, 12.5 Hz, 1 H), 3.73 (dt, J = 5.2, 11.0 Hz, 1 H), 3.32 (td, J = 1.9, 11.3 Hz, 1 H); ¹³C NMR (126MHz, CDCl₃) $\delta = 164.7$, 156.0, 137.8, 137.5, 135.6, 131.0, 130.0, 129.4, 128.4, 127.9, 122.6, 118.6, 114.8, 113.0, 112.6, 79.2, 77.1, 66.5, 43.2, 42.7. HRMS (ESI) calcd for $C_{20}H_{14}ClN_3O_3 [M+H]^+$: 380.0796, found: 386.0809; IR (KBr): v 3470, 2225, 1572, 1556, 1497, 1363, 1094, 1010, 827, 784, 736; 88% ee was determined by HPLC on ODH column, 10/90% 2-propanol/hexane, 1.0mL/min, UV 254nm, $t_{minor} = 18.3 \text{ min}, t_{major} = 20.8 \text{ min}.$

2-((S)-3-((R)-1-(4-chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile:

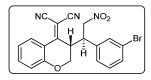
General experimental procedure II was followed to prepare the Michael addition product 3bd.



The desired product was obtained as white solid with 80% yield, mp: 177-179 °C; ¹H NMR (500MHz, CDCl₃) δ = 8.28 (dd, J = 1.6, 8.2 Hz, 1 H), 7.64 (ddd, J = 1.4, 7.2, 8.4 Hz, 1 H), 7.46 - 7.41 (m, 2 H), 7.29 (d, J = 8.5 Hz, 2 H), 7.21 - 7.15 (m, 1 H), 7.09 (dd, J = 0.9, 8.5 Hz, 1 H), 4.84 (dd, J = 10.7, 12.9 Hz, 1 H), 4.51 (dd, J = 5.2, 13.1 Hz, 1 H),

4.23 - 4.12 (m, 1 H), 4.12 - 4.03 (m, 1 H), 3.74 (dt, J = 5.0, 11.0 Hz, 1 H), 3.29 (td, J = 1.9, 11.4 Hz, 1 H). ¹³C NMR (125 MHz, CDCl₃) $\delta = 164.8$, 156.0, 137.8, 135.2, 135.1, 133.9, 129.9, 129.5, 127.9, 122.6, 118.6, 114.8, 113.1, 112.7, 79.1, 77.2, 66.5, 43.0. HRMS (ESI) calcd for C₂₀H₁₄ClN₃O₃ [M+H]⁺: 380.0796, found: 386.0802. IR (KBr): v 3468, 2235, 1580, 1560, 1499, 1365, 1097, 1015, 829, 784, 735 cm⁻¹; 93% ee was determined by HPLC on AS column, 15/85% 2-propanol/hexane, 1.0mL/min, UV 254nm, $t_{minor} = 29.5$ min, $t_{maior} = 32.3$ min.

2-((S)-3-((R)-1-(3-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile:



General experimental procedure **II** was followed to prepare the Michael addition product **3be.** The desired product was obtained as white solid with 79% yield, mp: 182-185 °C; ¹H NMR (500MHz, CDCl₃) δ = 8.28 (dd, J = 1.6, 8.2 Hz, 1 H), 7.64 (ddd, J = 1.4, 7.2, 8.4 Hz, 1 H), 7.55 (qd, J = 1.1, 7.9 Hz, 1 H), 7.48 (t, J = 1.7 Hz, 1 H), 7.37 - 7.32 (m, 1

H), 7.30 (t, J = 1.4 Hz, 1 H), 7.18 (ddd, J = 0.9, 7.3, 8.2 Hz, 1 H), 7.10 (dd, J = 1.1, 8.4 Hz, 1 H), 4.84 (dd, J = 10.6, 13.1 Hz, 1 H), 4.52 (dd, J = 5.0, 13.2 Hz, 1 H), 4.18 (dd, J = 2.4, 12.5 Hz, 1 H), 4.09 (dd, J = 1.7, 12.5 Hz, 1 H), 3.72 (dt, J = 5.2, 11.0 Hz, 1 H), 3.32 (td, J = 2.0, 11.4 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) $\delta = 164.7$, 156.0, 137.8, 137.8, 132.4, 131.3, 131.2, 127.9, 126.6, 123.7, 122.6, 118.6, 114.8, 113.0, 112.6, 79.2, 77.1, 66.5, 43.1, 42. HRMS (ESI)

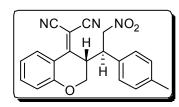
calcd for $C_{20}H_{14}BrN_3O_3$ [M+Na]⁺: 446.0110, found: 446.0121; IR (KBr): 3468, 2928, 2232, 1585, 1561, 1494, 1437, 1375, 1094, 1015, 830, 768, 738, 641; 87% ee was determined by HPLC on ODH column, 20/80% 2-propanol/hexane, 1.0mL/min, UV 254nm, $t_{minor} = 17.6$ min, $t_{maior} = 21.9$ min.

2-((S)-3-((R)-1-(4-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile

General experimental procedure **II** was followed to prepare the Michael addition product **3bf.** The desired product was obtained as white solid with 83% yield, mp: 178-180 °C; 1 H NMR (500MHz, CDCl₃) δ = 8.28 (dd, J = 1.3, 8.2 Hz, 1 H), 7.64 (ddd, J = 1.6, 7.1, 8.4 Hz, 1 H), 7.62 - 7.56 (m, 2 H), 7.26 - 7.20 (m, 2 H), 7.20 - 7.15 (m, 1 H), 7.09 (dd, J =

0.9, 8.5 Hz, 1 H), 4.84 (dd, J = 10.7, 12.9 Hz, 1 H), 4.51 (dd, J = 5.0, 12.9 Hz, 1 H), 4.16 (dd, J = 2.4, 12.5 Hz, 1 H), 4.12 - 4.05 (m, 1 H), 3.73 (dt, J = 5.0, 11.0 Hz, 1 H), 3.29 (td, J = 2.0, 11.4 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) $\delta = 164.8$, 156.0, 137.8, 134.4, 132.9, 129.7, 127.9, 123.3, 122.6, 118.6, 114.8, 113.1, 112.7, 79.1, 77.2, 66.5, 43.0, 42.7. HRMS (ESI) calcd for $C_{20}H_{14}BrN_3O_3[M+Na]^+$: 446.0110, found: 446.0121; IR (KBr): v 3466, 2929, 2234, 1587, 1564, 1497, 1439, 1379, 1095, 1017, 832, 770, 740, 641 cm⁻¹; 86% ee was determined by HPLC on ODH column, 10/90% 2-propanol/hexane, 1.0mL/min, UV 254nm, $t_{minor} = 12.6$ min, $t_{major} = 18.6$ min.

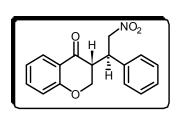
2-(2-(1-(4-Methylphenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile.



General experimental procedure **II** was followed to prepare the Michael addition product **3bg.** The desired product was obtained as white solid with 81% yield mp: 183-185°C; ¹H NMR (500MHz, CDCl₃) δ (ppm) 8.26 (dd, J = 1.0, 8.0 Hz, 1H), 7.63 (td, J = 7.5, 0.5 Hz, 1H), 7.29-7.26 (m, 4H), 7.17 (td, J = 1, 7.0 Hz, 1H), 7.07 (dd, J = 1.0, 8.5 Hz, 1H), 4.84 (dd, J = 2.5, 13.0 Hz, 1H), 4.49 (dd, J = 5.5, 12.5Hz, 1H), 4.12- 4.11 (m, 2H), 3.69 (dt, J = 3, 11.5 Hz,

1H), 3.30 (dd, J = 2, 13.5Hz, 1H; ¹³C NMR (125 MHz, CDCl₃) δ (ppm) 174.4, 166.0, 156.1, 139. 9, 137.7, 132.2, 131.7, 130.3, 127.9, 122.4, 114.9, 78.9, 77.3, 66.6, 43.3, 43.1, 21.1; HRMS (ESI) calcd for C₂₁H₁₇N₃O₃ [M+H]⁺: 360.1342, found: 360.1331; IR (KBr): 3468, 2230, 1574, 1559, 1515, 1378, 1260, 1180, 1037, 828, 775, 751 cm-1; 90% ee was determined by HPLC on ADH column, 15/85% 2-propanol/hexane, 1.0mL/min, UV 254nm, $t_{minor} = 21.5$ min, $t_{major} = 26.4$ min.

(R)-3-((R)-2-nitro-1-phenylethyl)chroman-4-one:



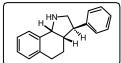
To a stirred solution of 3ba (1 equiv) in acetone and water as solvent, $KMnO_4$ (2 equiv) was added. The solution was stirred at ambient temperature for 4h. After the reaction was completed (monitored by TLC), the resulting mixture was concentrated under reduced pressure and the residue was purified through column chromatography on silica gel to give the product 7b. The desired product 7b was obtained as white solid with 67% yield; mp: 155-

157°C; ¹H NMR (500MHz, CDCl₃) δ = 7.95 (dd, J = 1.9, 7.9 Hz, 1 H), 7.56 (ddd, J = 1.7, 7.1, 8.5 Hz, 1 H), 7.44 - 7.40 (m, 2 H), 7.38 - 7.34 (m, 1 H), 7.33 - 7.30 (m, 2 H), 7.13 - 7.08 (m, 1 H), 7.04 - 7.00 (m, 1 H), 4.96 (dd, J = 5.0, 13.2 Hz, 1 H), 4.82 (dd, J = 10.6, 13.4 Hz, 1 H), 4.28

(dd, J = 3.5, 11.7 Hz, 1 H), 4.04 (dd, J = 5.2, 11.8 Hz, 1 H), 3.95 (dt, J = 4.7, 10.9 Hz, 1 H), 2.84 - 2.78 (m, 1 H); ¹³C NMR (125 MHz, CDCl₃) $\delta = 192.8$, 161.4, 136.7, 136.3, 129.4, 128.5, 128.0, 127.7, 122.0, 120.0, 117.9, 78.5, 68.7, 48.6, 41.2; HRMS (ESI) calcd for $C_{17}H_{15}NO_4$ [M+H]⁺: 298.1074, found: 298.1087; IR (KBr): 3467, 2228, 1779, 1575, 1560, 1525, 1381, 1265, 1182, 1034, 820, 770 cm⁻¹;

(3R,3aS,9bR)-3-phenyl-2,3,3a,4,5,9b-hexahydro-1H-benzo[g]indole:

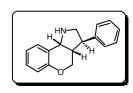
To a stirred solution of 7a (1 equiv) in acetic acid as solvent, Zn dust (10 equiv) was added. The



solution was stirred at ambient temperature for 8h. After the reaction was completed (monitored by TLC), the resulting mixture was concentrated under reduced pressure and the residue was purified through column chromatography on silica gel to give the product 8a. The desired product

was obtained as pasty solid with 61% yield; ¹H NMR (500MHz, CDCl₃) δ = 7.47 (d, J = 7.3 Hz, 1 H), 7.37 - 7.29 (m, 4 H), 7.25 - 7.10 (m, 4 H), 4.45 (d, J = 8.2 Hz, 1 H), 3.49 (dd, J = 6.8, 10.6 Hz, 1 H), 3.09 - 3.04 (m, 1 H), 3.04 - 2.98 (m, 1 H), 2.84 - 2.77 (m, 1 H), 2.64 (ddd, J = 4.1, 9.1, 15.8 Hz, 1 H), 2.55 - 2.45 (m, 1 H), 1.95 - 1.88 (m, 1 H), 1.60 (dtd, J = 4.1, 8.8, 13.2 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ = 144.5, 138.4, 138.2, 129.7, 128.9, 128.4, 127.9, 126.7, 126.5, 126.4, 59.7, 55.1, 52.6, 46.9, 28.3, 26.7; HRMS (ESI) calcd for C₁₈H₁₉N [M+Na]⁺: 298.1074, found: 298.1087; IR (KBr): 3625, 3650, 2230, 1625, 1556, 1378, 1271, 1185, 1037, 838, 779 cm⁻¹;85% ee was determined by HPLC on phenomenex Amylose 2 column, 20/80% 2-propanol/hexane, 1.0mL/min, UV 254nm, t_{minor} = 19.6 min, t_{major} = 22.3 min.

(3R,3aS,9bR)-3-phenyl-1,2,3,3a,4,9b-hexahydrochromeno[4,3-b]pyrrole:



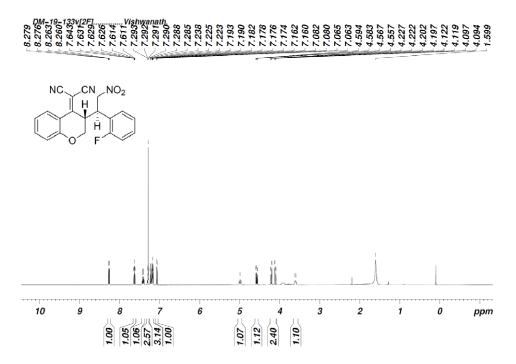
To a stirred solution of **7b** (1 equiv) in acetic acid as solvent, Zn dust (10 equiv) was added. The solution was stirred at ambient temperature for 8h. After the reaction was completed (monitored by TLC), the resulting mixture was concentrated under reduced pressure and the residue was purified through column chromatography on silica gel to give the product **8b**. The desired product **8b** was obtained as pasty solid with 64% yield; ¹H

NMR (500MHz, DMSO-d₆) δ = 7.40 (dd, J = 1.6, 7.6 Hz, 1 H), 7.36 - 7.30 (m, 4 H), 7.25 - 7.20 (m, 1 H), 7.18 - 7.12 (m, 1 H), 6.94 (dt, J = 1.3, 7.4 Hz, 1 H), 6.84 (dd, J = 1.3, 8.2 Hz, 1 H), 4.23 - 4.12 (m, 2 H), 3.67 (dd, J = 9.8, 10.7 Hz, 1 H), 3.31 (dd, J = 7.9, 10.7 Hz, 1 H), 2.88 (d, J = 5.4 Hz, 1 H), 2.77 (dd, J = 8.7, 10.6 Hz, 1 H), 2.43 (tdd, J = 5.1, 6.9, 9.8 Hz, 1 H); ¹³C NMR (125 MHz, DMSO-d₆) δ = 155.1, 144.4, 131.1, 129.0, 128.3, 128.0, 126.7, 125.1, 121.2, 116.9, 66.0, 56.3, 55.3, 48.3, 45.8; HRMS (ESI) calcd for $C_{17}H_{17}NO$ [M+H]⁺: 252.1383, found: 252.1370; IR (KBr): 3628, 3655, 2235, 1628, 1558, 1374, 1275, 1182, 1039, 845, 740 cm⁻¹; 75% ee was determined by HPLC on ODH column, 30/70% 2-propanol/hexane, 1.0mL/min, UV 254nm, t_{minor} = 49.6 min, t_{major} = 59.0 min.

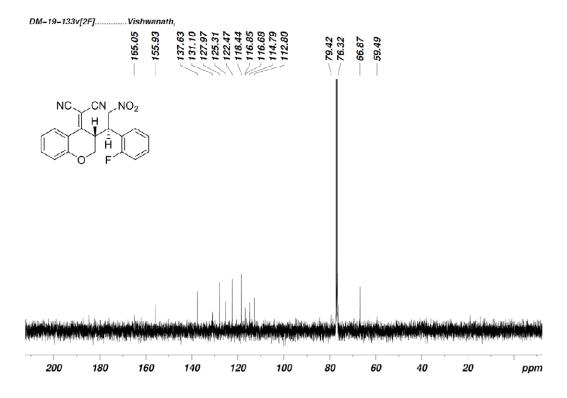
References:

- 1) D. Xue, Y.-C. Chen, Q.-W. Wang, L.-F. Cun, J. Zhu, J.-G. Deng, Org. Lett. 2005, 7, 5293-5296;
- 2) T. B. Poulsen, M. Bell, K. A. Jorgensen, Org. Biomol. Chem. 2006, 4, 63-70.
- 3) Zhou, L.-H.; Wang, N.; Chen, G.-N.; Yang, Q.; Yang, S.-Y.; Zhang, W.; Zhang, Y.; Yu, X.-Q. Journal of Molecular Catalysis B: Enzymatic 2014, 109, 170-177

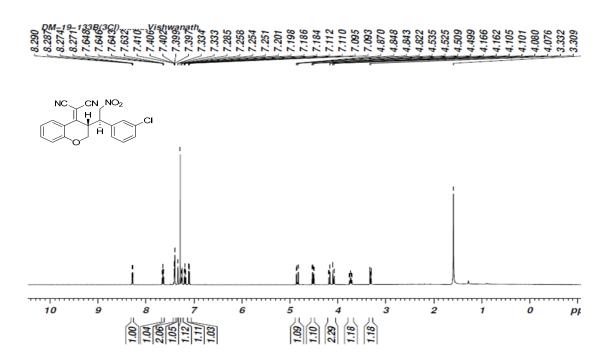
$^1\mathrm{H}$ NMR of 2-(2-(1-(2-fluorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile



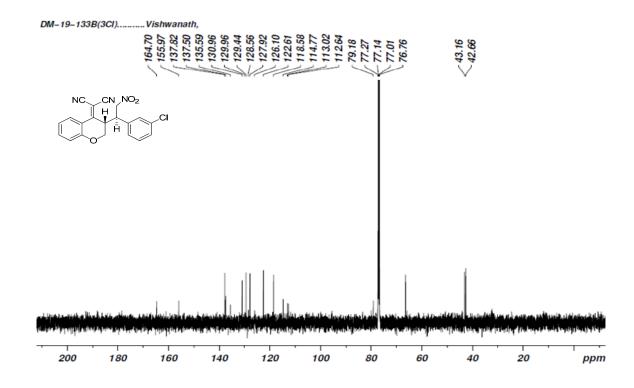
 $^{\rm 13}{\rm C~NMR}$ of 2-(2-(1-(4-fluorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile



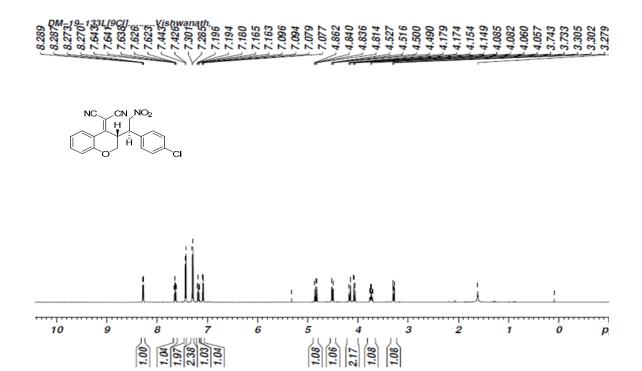
¹H NMR of 2-(3-(1-(3-Chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.



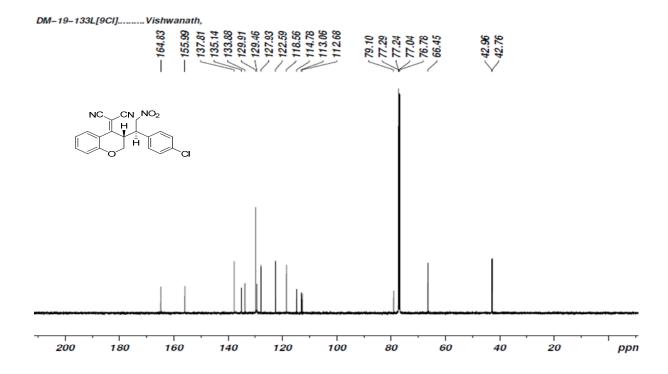
¹³C NMR of 2-(3-(1-(3-Chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.



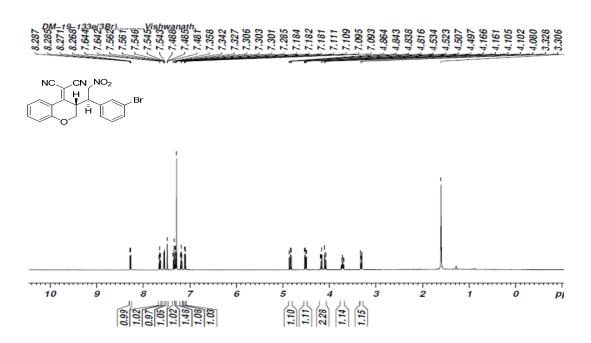
¹H NMR of 2-(3-(1-(4-chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.



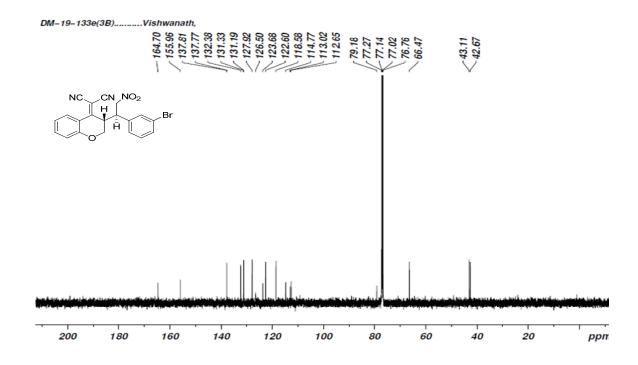
¹³C NMR of 2-(3-(1-(4-chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.



¹H NMR of 2-(3-(1-(3-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.

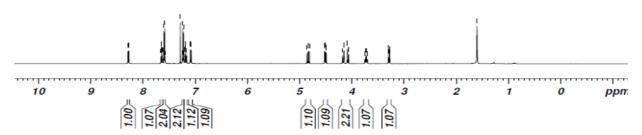


¹³C NMR of 2-(3-(1-(3-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.

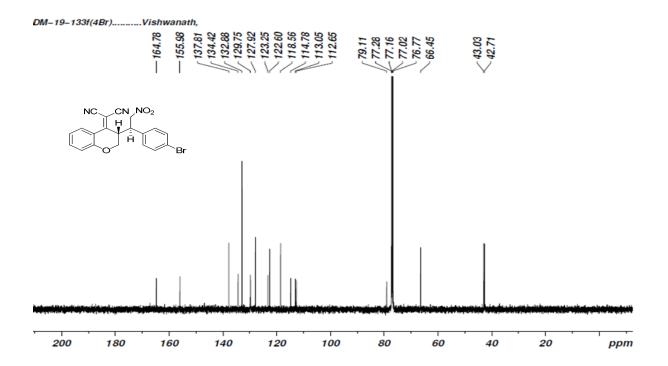


¹H NMR of 2-(3-(1-(4-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.

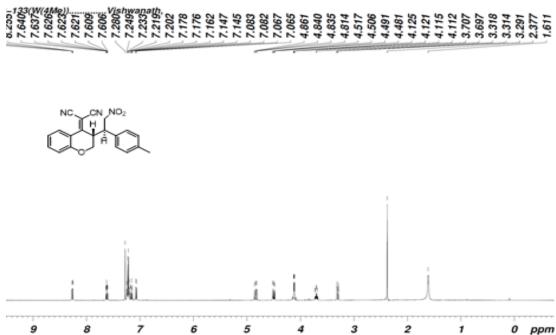




¹³C NMR of 2-(3-(1-(4-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.

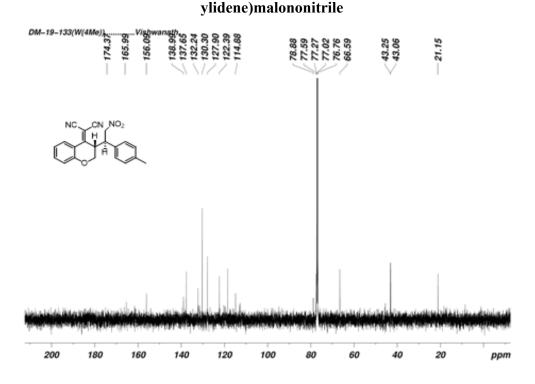


 1 H NMR of 2-(2-(1-(4-Methylphenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile

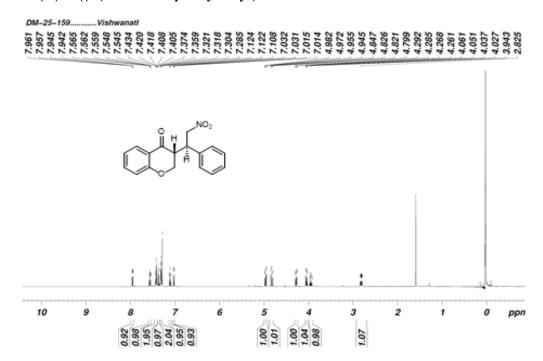


 $^{13}\mathrm{C}$ NMR of 2-(2-(1-(4-Methylphenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile

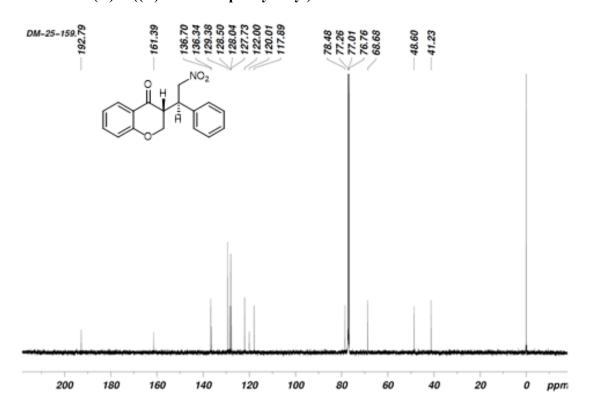
1.11



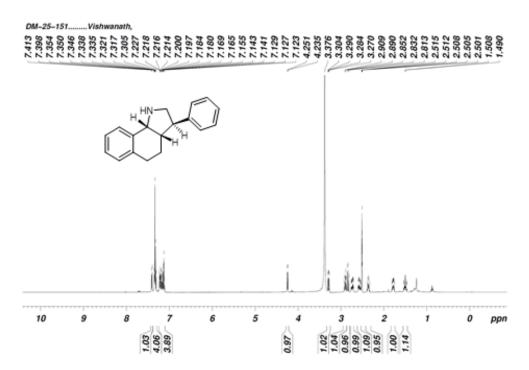
¹H NMR of (R)-3-((R)-2-nitro-1-phenylethyl)chroman-4-one.



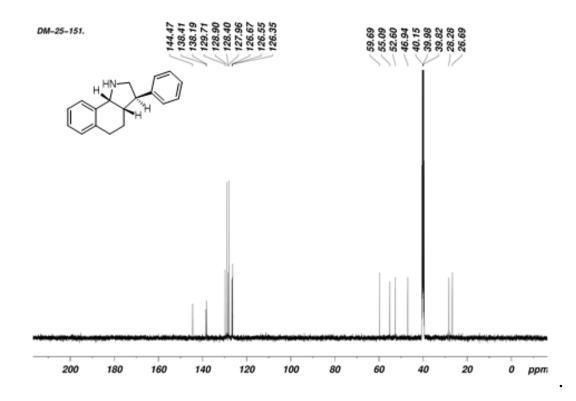
¹³C NMR of (R)-3-((R)-2-nitro-1-phenylethyl)chroman-4-one



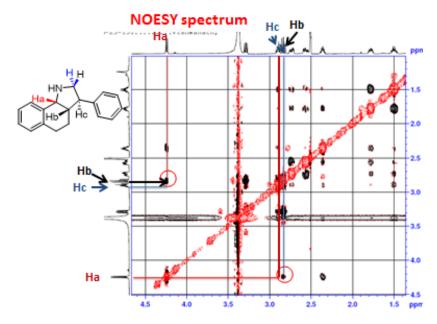
¹H NMR of (3R,3aS,9bR)-3-phenyl-2,3,3a,4,5,9b-hexahydro-1H-benzo[g]indole.



¹³C NMR of (3R,3aS,9bR)-3-phenyl-2,3,3a,4,5,9b-hexahydro-1H-benzo[g]indole.

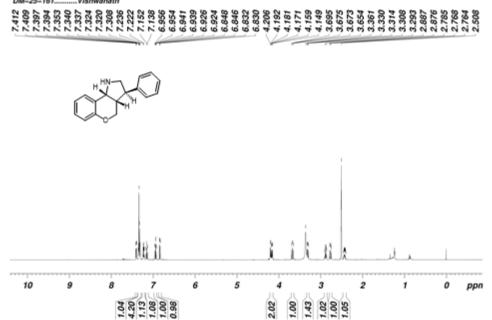


NOESY spectrum of (3R,3aS,9bR)-3-phenyl-2,3,3a,4,5,9b-hexahydro-1H-benzo[g]indole.

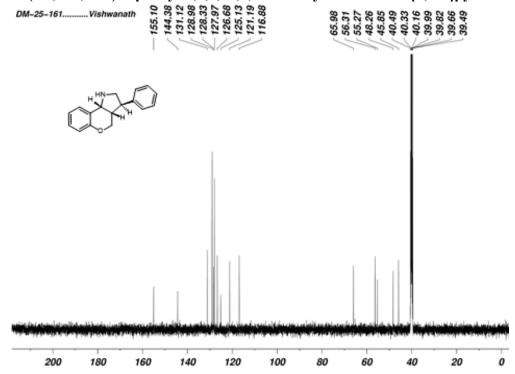


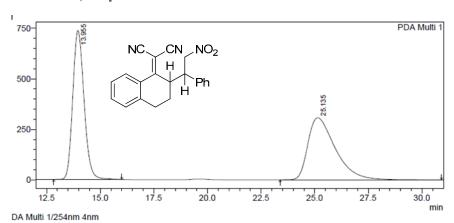
NOESY spectum shows no spatial interactions between ${\it Ha}$ and ${\it Hc}$, which indicates those protons are in "anti" configuration in cycl ised product.

¹H NMR of (3R.3aS.9bR)-3-phenvl-1.2.3.3a.4.9b-hexahvdrochromeno[4.3-blpvrrole



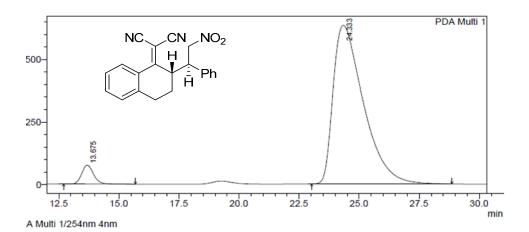
¹³C NMR of (3R,3aS,9bR)-3-phenyl-1,2,3,3a,4,9b-hexahydrochromeno[4,3-b]pyrrole.





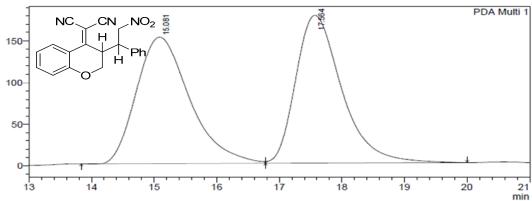
PeakTable

PDA Ch1 2	54nm 4nm				
Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.955	26947195	735911	49.599	70.545
2	25.135	27383159	307271	50.401	29.455
Total		54330354	1043181	100.000	100.000



PeakTable

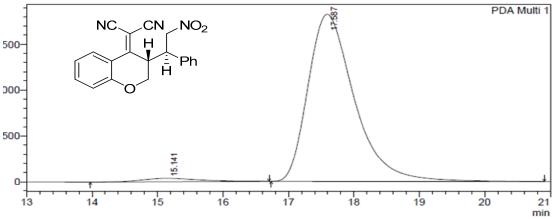
PDA Chi 2	254nm 4nm				
Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.675	2569671	75341	4.419	10.609
2	24.333	55575036	634836	95.581	89.391
Total		58144707	710176	100.000	100.000



)A Multi 1/220nm 4nm

PeakTable

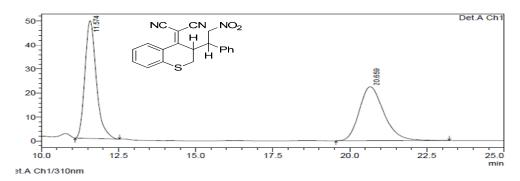
PDA Cn1 220nm 4nm						
	Peak#	Ret. Time	Area	Height	Area %	Height %
- 1	1	15.081	9015261	151860	49.881	46.087
	2	17.564	9058285	177646	50.119	53.913
1	Total		18073545	329505	100.000	100.000



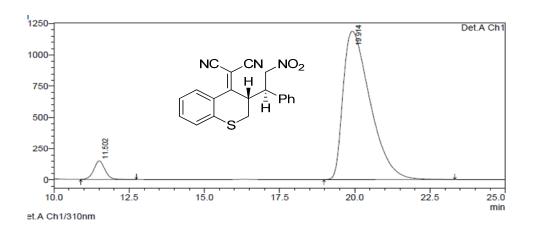
A Multi 1/220nm 4nm

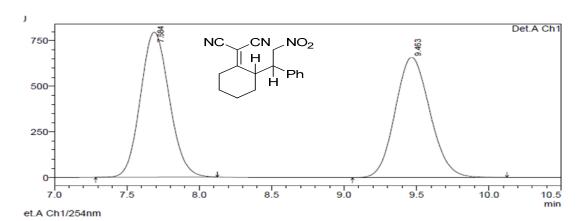
PeakTable

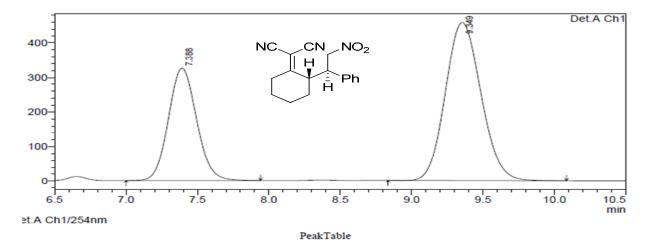
PDA Ch1 220nm 4nm						
	Peak#	Ret. Time	Area	Height	Area %	Height %
Ī	1	15.141	2168839	36067	2.366	1.938
	2	17.587	89482351	1824522	97.634	98.062
	Total		91651190	1860589	100.000	100.000



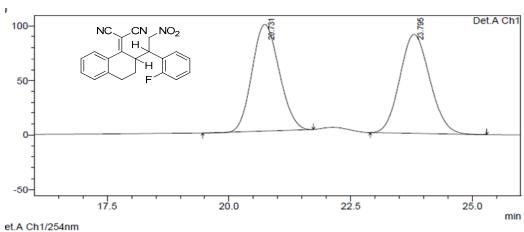
PeakTable								
Detector A Ch1 310nm								
Peak#	Ret. Time	Area	Height	Area %				
1	11.574	1261592	49068	50.309				
2	20.659	1246085	22342	49.691				
Total		2507677	71411	100.000				
31	% 8.713 1.287 0.000							







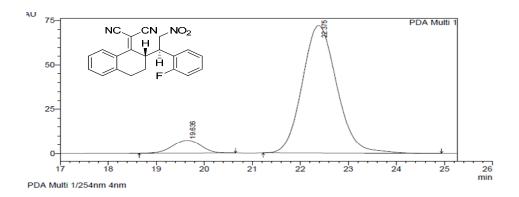
Detector A	Ch1 254nm				
Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.388	4470846	325709	35.990	41.577
2	9.349	7951607	457681	64.010	58.423
Total		12422452	783390	100.000	100.000



et.A CITI/204IIIII

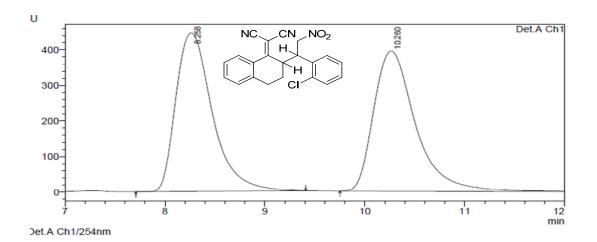
PeakTable

	Detector A Ch1 254nm					
	Peak#	Ret. Time	Area	Height	Area %	Height %
	1	20.731	3785151	97609	49.380	51.954
ı	2	23.795	3880135	90265	50.620	48.046
Ī	Total		7665286	187874	100.000	100.000



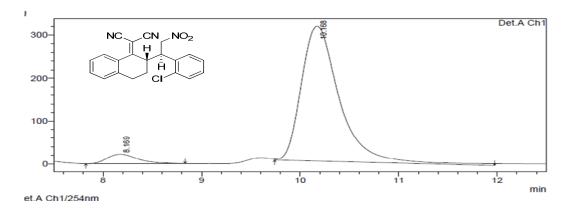
PeakTable

PDA CIII 2					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.636	289704	7079	7.480	8.968
2	22.375	3583401	71856	92.520	91.032
Total		3873105	78936	100.000	100.000



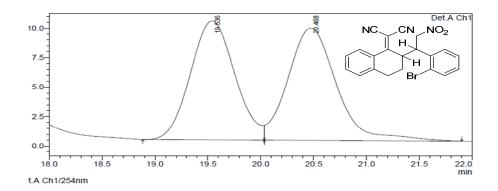
PeakTable

Detector A	Cn1 234nm				
Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.258	11288799	446150	49.533	53.127
2	10.260	11501444	393635	50.467	46.873
Total		22790243	839784	100.000	100.000



Peak Table										
		1.	ь	2	т	r	2	_	D	
	c	м	v	a		ъ.	GL.	c		

	CH1 234IIII				
Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.169	464727	21637	5.208	6.442
2	10.168	8458352	314233	94.792	93.558
3	14.933	0	0	0.000	0.000
Total		8923080	335871	100.000	100.000



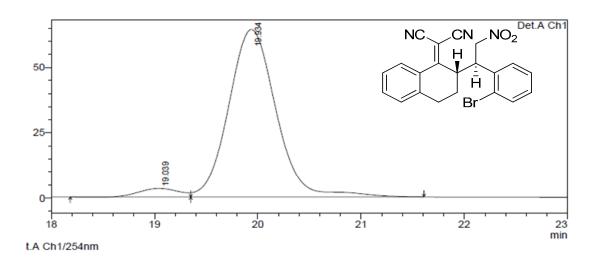
 PeakTable

 Detector A Ch1 254nm
 Peak# Ret. Time
 Area
 Height
 Area %
 Height %

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 19.536
 294078
 10079
 49.235
 51.411

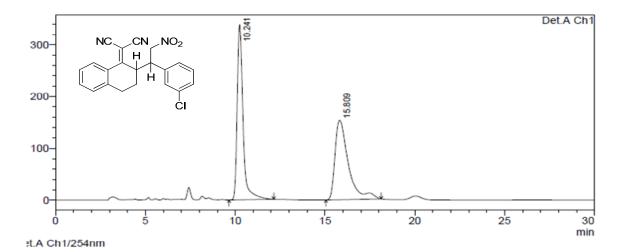
 2
 2 20.468
 303213
 9526
 50.765
 48.589

 Total
 597291
 19604
 100.000
 100.000



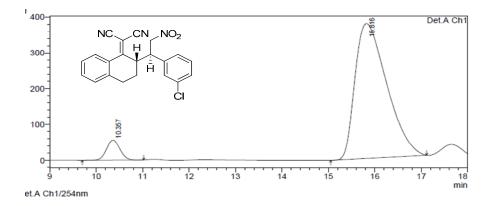
PeakTable |

Detector A Ch1 254nm						
	Peak#	Ret. Time	Area	Height	Area %	Height %
	1	19.039	83939	3156	3.965	4.696
ĺ	2	19.934	2033259	64049	96.035	95.304
	Total		2117199	67205	100.000	100.000



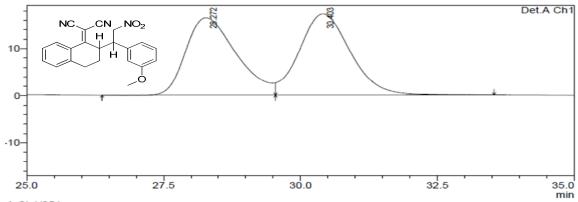
PeakTable

Detector A Ch1 254nm						
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	10.241	7678770	338919	49.292	68.827	
2	15.809	7899399	153502	50.708	31.173	
Total		15578168	492421	100.000	100.000	



PeakTable

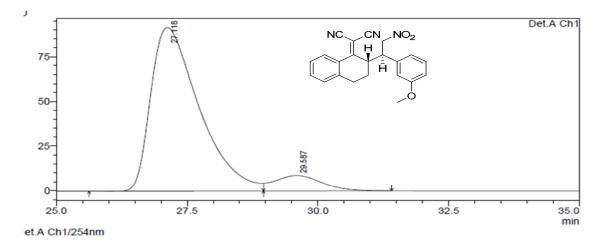
Detector.	Defector A Ciri 234iiii						
Peak#	Ret. Time	Area	Height	Area %	Height %		
	1 10.357	1097625	55457	5.789	12.821		
	2 15.816	17861565	377079	94.211	87.179		
Tot	al	18959190	432537	100.000	100.000		



A Ch1/254nm

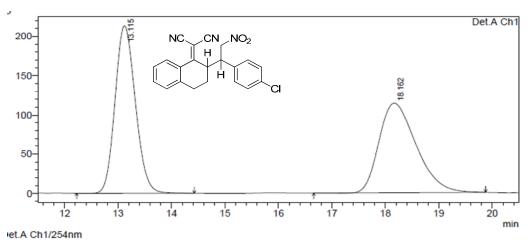
PeakTable

D	Detector A Ch1 254nm						
	Peak#	Ret. Time	Area	Height	Area %	Height %	
	1	28.272	1050746	16429	48.566	48.791	
	2	30.403	1112818	17242	51.434	51.209	
	Total		2163564	33671	100.000	100.000	



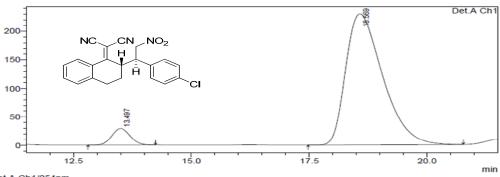
PeakTable

Detector A Ch1 254nm								
Peak#	Ret. Time	Area	Height	Area %	Height %			
1	27.118	5895994	91631	91.538	91.357			
2	29.587	545072	8669	8.462	8.643			
Total		6441065	100300	100.000	100.000			



PeakTable

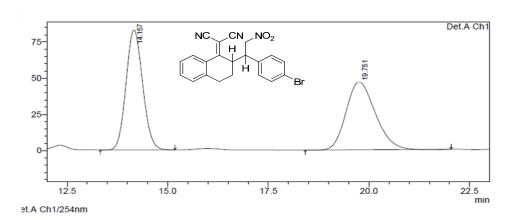
Detector A	Detector A Ch1 254nm							
Peak#	Ret. Time	Area	Height	Area %	Height %			
1	13.115	5624880	212978	50.629	65.124			
2	18.162	5485166	114057	49.371	34.876			
Total		11110045	327034	100.000	100.000			



et.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.497	769089	28600	5.796	11.091
2	18.569	12499451	229261	94.204	88.909
Total		13268540	257861	100.000	100.000



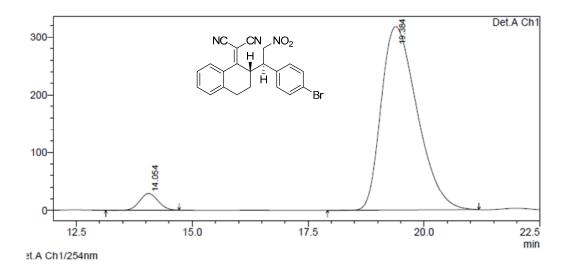
Height % 64.002 35.998 100.000

 PeakTable

 Detector A Ch1 254nm

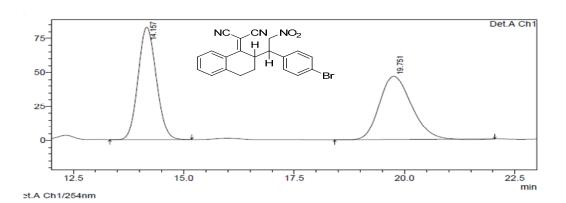
 Peak#
 Ret. Time
 Area
 Height
 Area %

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 82770
 50.399



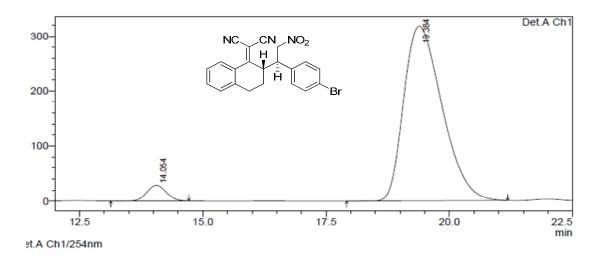
PeakTable

Detector A	Detector A Cn1 254mm						
Peak#	Ret. Time	Area	Height	Area %	Height %		
1	14.054	781132	28397	4.364	8.213		
2	19.384	17117417	317366	95.636	91.787		
Total		17898550	345763	100.000	100.000		



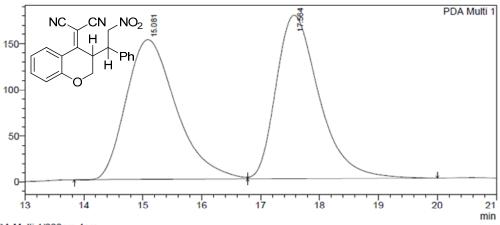
PeakTable

Detector A Ch1 254nm						
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	14.157	2371480		50.399	64.002	
2	19.751		46554	49.601	35.998	
Total		4705453	129324	100.000	100.000	



PeakTable

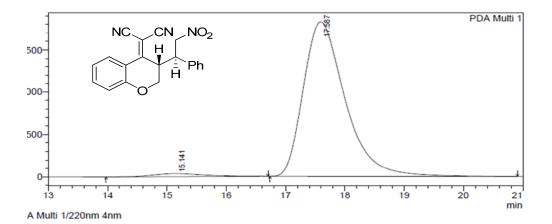
	Detector A Ch1 254nm							
	Peak#	Ret. Time	Area	Height	Area %	Height %		
ı	1	14.054	781132	28397	4.364	8.213		
ı	2	19.384	17117417	317366	95.636	91.787		
ı	Total		17898550	345763	100,000	100,000		



OA Multi 1/220nm 4nm

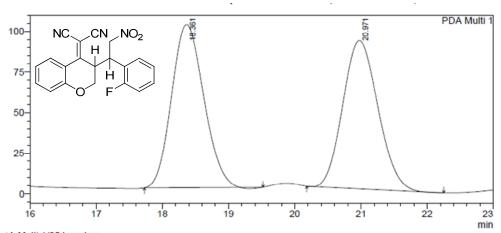
PeakTable

	PDA Ch1 220nm 4nm						
	Peak#	Ret. Time	Area	Height	Area %	Height %	
	1	15.081	9015261	151860	49.881	46.087	
1	2	17.564	9058285	177646	50.119	53.913	
Ì	Total		18073545	329505	100.000	100.000	



PeakTable

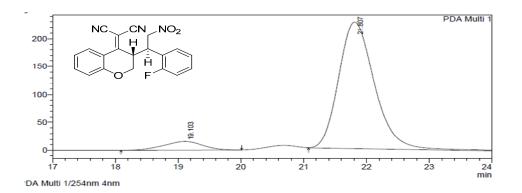
PDA Chi 2	PDA Chi 220nm 4nm					
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	15.141	2168839	36067	2.366	1.938	
2	17.587	89482351	1824522	97.634	98.062	
Total		91651190	1860589	100.000	100.000	



A Multi 1/254nm 4nm

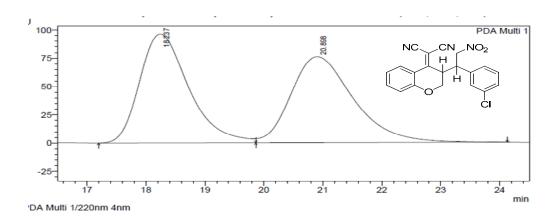
PeakTable

]	PDA Ch1 254nm 4nm							
[Peak#	Ret. Time	Area	Height	Area %	Height %		
	1	18.361	3382550	100386	50.075	52.415		
	2	20.971	3372374	91134	49.925	47.585		
-[Total		6754924	191519	100.000	100.000		

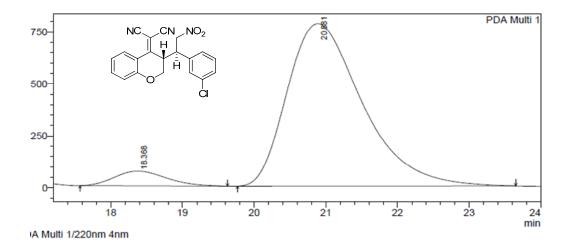


PeakTable

PDA Ch1 254nm 4nm							
Peak#	Ret. Time	Area	Height	Area %	Height %		
1	19.103	662258	15482	6.821	6.384		
2	21.807	9046242	227034	93.179	93.616		
Total		9708500	242516	100.000	100.000		



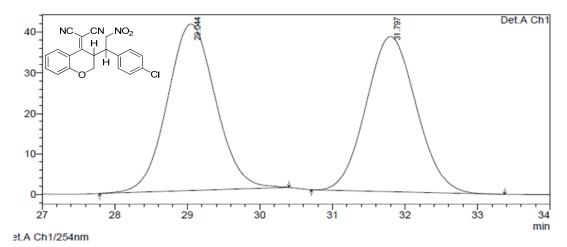
		r	Cakiable		
PDA Ch1 2	20nm 4nm				
Peak#	Ret. Time	Area	Height	Area %	Height %
1	18.237	5589148	96384	49.918	55.944
2	20.898	5607447	75903	50.082	44.056
Total		11106505	172287	100 000	100 000



PDA Ch1 220nm 4nm

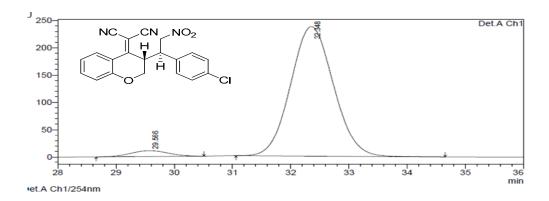
| Peak# | Ret. Time | 18.368 | 20.881 Area % 5.814 94.186 100.000 Area 3519944 57027713 60547657 Height 69919 782674 852593

PeakTable



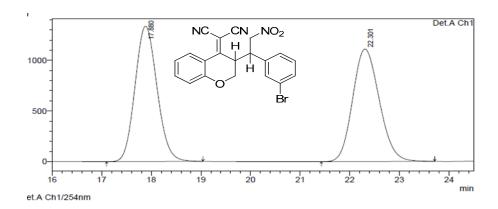
PeakTable

Detector A					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	29.044	1893763	41025	50.683	51.835
2	31.797	1842691	38121	49.317	48.165
Total		3736454	79146	100.000	100.000



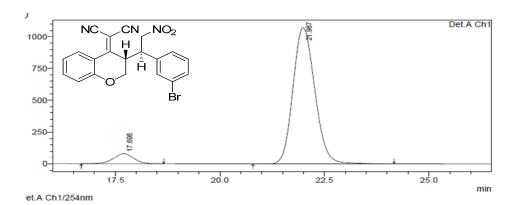
PeakTable

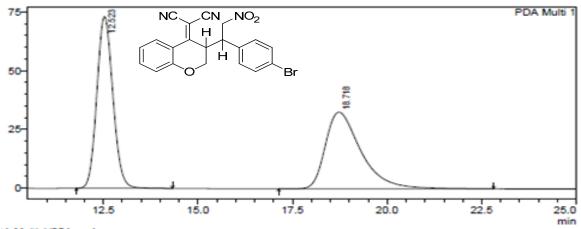
Peak#	Ret. Time	Area	Height	Area %	Height %
1	29.566	475140	10639	3.767	4.294
2	32.348	12138613	237148	96.233	95.706
Total		12613754	247787	100.000	100.000



PeakTable

Detector A					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	17.880	41183161	1334860	49.888	54.610
2	22.301	41368840	1109496	50.112	45.390
Total		82552002	2444356	100.000	100.000

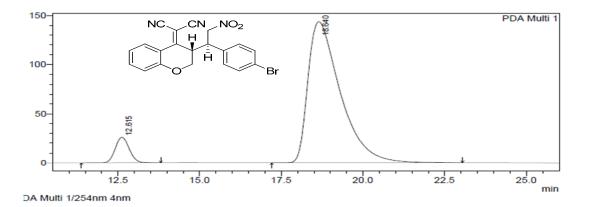




IA Multi 1/254nm 4nm

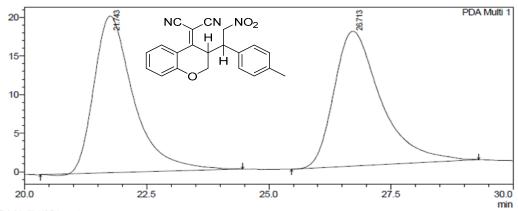
PeakTable

PDA Chl :					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	12.523	2154083	73118	50.258	69.154
2	18.718	2131933	32614	49.742	30.846
Total		4286016	105732	100.000	100.000



PeakTable

PDA Ch1 2					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	12.615	774877	25984	7.263	15.334
2	18.640	9894663	143472	92.737	84.666
Total		10669539	169455	100.000	100.000

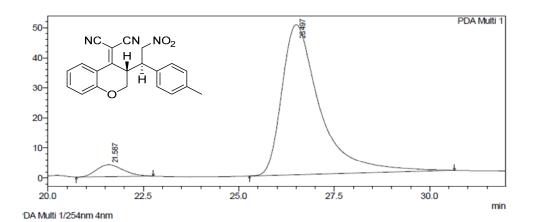


DA Multi 1/254nm 4nm

PeakTable

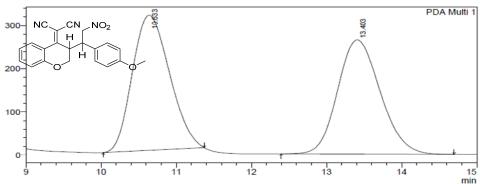
PDA Ch1 25	PDA Ch1 254nm 4nm								
Peak#	Ret. Time	Area	Height	Area %					
1	21.743	1146070	20286	49.465					
2	26.713	1170878	17472	50.535					
Total		2316948	37758	100.000					

Height %
53.727
46.273
100.000



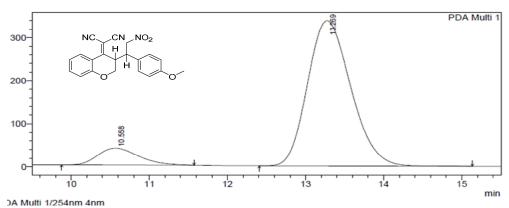
PeakTable

PDA Ch1 254nm 4nm						
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	21.587	190903	3889	5.209	7.228	
2	26.497	3473651	49914	94.791	92.772	
Total		3664554	53803	100.000	100.000	



OA Multi 1/254nm 4nm

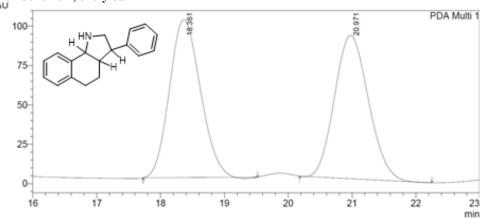
		F	Cartable		
PDA Ch1 2	54nm 4nm				
Peak#	Ret. Time	Area	Height	Area %	Height %
1	10.633	10796471	313123	50.829	54.145
2	13.403	10444326	265186	49.171	45.855
Total		21240797	578309	100.000	100.000



PDA Ch1 254nm 4nm PeakTable

Total Control Control						
j	Peak#	Ret. Time	Area	Height	Area %	Height %
1	1	10.558	1416199	39060	9.831	10.357
	2	13.269	12988798	338071	90.169	89.643
ı	Total		14404997	377131	100.000	100.000

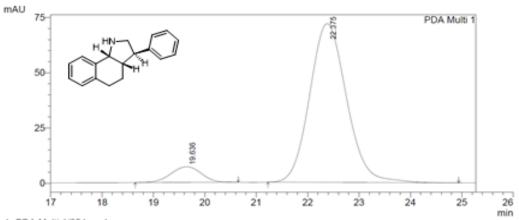
HPLC profile for Scheme 2, entry 8a



1 PDA Multi 1/254nm 4nm

PeakTable

DA Ch1 254nm 4nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	18.361	3382550	100386	50.075	52.415
2	20.971	3372374	91134	49.925	47.585
Total		6754924	191519	100.000	100.000

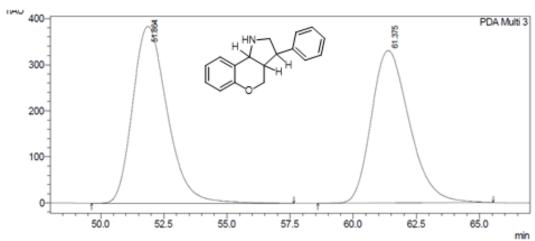


1 PDA Multi 1/254nm 4nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.636	289704	7079	7.480	8.968
2	22.375	3583401	71856	92.520	91.032
Total		3873105	78936	100.000	100.000

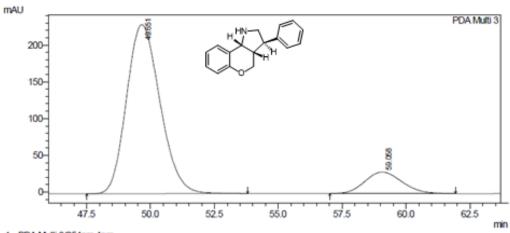
HPLC profile for Scheme 2, entry 8b



1 PDA Multi 3/254nm 4nm

PeakTable

PDA Ch3 254nm 4nm						
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	51.864	36557083	383877	51.339	53.730	
2	61.375	34649864	330580	48.661	46.270	
Total		71206947	714457	100.000	100.000	



1 PDA Multi 3/254nm 4nm

PeakTable

PDA Ch3 254nm 4nm						
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	49.651	20982936	230042	87.569	88.691	
2	59.058	2978791	29334	12.431	11.309	
Total		23961727	259376	100,000	100,000	