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# Construction of polycyclic fused pyrrolidines with three contiguous stereocentres via Michael addition of vinyl malononitriles with nitrostyrenes using $L$ Proline derived thiourea 

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## I. General Remarks:

All reactions were carried out in an oven dried flask. Solvents used for reactions and column chromatography were commercial grade and distilled prior to use. Toluene and THF were dried over sodium/benzophenone, $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ and $\mathrm{CHCl}_{3}$ over $\mathrm{CaH}_{2}$. Solvents for HPLC bought as analytical grade and used without further purification. TLC was performed on precoated Merck silica gel aluminium plates with $60_{\mathrm{F}} 254$ indicator, visualised by irradiation with UV light. Column chromatography was performed using silica gel Merck 60-100 mesh. ${ }^{1} \mathrm{H}$-NMR and ${ }^{13} \mathrm{C}$-NMR were recorded on a Bruker AV 500 MHz using $\mathrm{CD}_{3} \mathrm{OD}-\mathrm{d}_{4}$ and $\mathrm{CDCl}_{3}$ as solvent and multiplicity indicated as follows: $s$ (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (doublet of doublet), dt (doublet of triplet) bs (broad singlet). Coupling constants J were reported in Hertz. High resolution mass spectra were obtained by ESI using Thermo scientific Orbitrap Elite mass spectrometer. IR spectra were recorded on a Perkin Elmer FT/IR-420 spectrometer and are reported in terms of frequency of absorption $\left(\mathrm{cm}^{-1}\right)$. The enantiomeric excess is obtained by HPLC analysis using a chiral stationary phase column (CHIRALPAK ADH, CHIRALCELL OD-H and Phenomenex Amylose-2. All the physical and spectroscopic data of 3a-d, 3aa- ah, 3ba, 3bh and $\mathbf{7 a}$ were in complete agreement with the reported literature.

## II. General procedure for asymmetric vinylogous Michael addition of vinyl malononitriles to nitrostyrene



To a stirred solution of $4(0.92 \mathrm{mg}, 0.002 \mathrm{mmol}, 5 \mathrm{~mol} \%)$ and vinyl malanonitriles $\mathbf{1}(0.1$ $\mathrm{mmol})$ in Toluene ( 1 mL ), nitrostyrene $2(0.12 \mathrm{mmol})$ was added. The solution was stirred at ambient temperature for mentioned days. After the reaction was completed (monitored by TLC), the resulting mixture was concentrated under reduced pressure and the residue was purified through column chromatography on silica gel to give the product 3.

2-((S)-2-((R)-2-nitro-1-phenylethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile 3a:

General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $28.5 \mathrm{mg}(0.083 \mathrm{mmol})$ with $83 \%$ yield; $91 \%$ ee determined by HPLC on AS column, 30\% 2-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}$, UV 254 nm , $\mathrm{t}_{\text {minor }}=13.3$ $\mathrm{min}, \mathrm{t}_{\text {major }}=20.4 \mathrm{~min}$.

## 2-((S)-3-((R)-2-nitro-1-phenylethyl)chroman-4 ylidene)malononitrile 3b:

General experimental procedure I was followed to prepare the Michael addtion product 3b. The desired product was obtained as white solid $27.9 \mathrm{mg}(0.081 \mathrm{mmol})$ with $81 \%$ yield; $95 \%$ ee was determined by HPLC on AS column, $30 \%$ 2-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}$, UV $254 \mathrm{~nm}, \mathrm{t}_{\text {minor }}=$ $15.14 \mathrm{~min}, \mathrm{t}_{\text {major }}=17.3 \mathrm{mmin} ;[\alpha]_{\mathrm{D}}{ }^{25}-172.3^{\circ}\left(c 0.15, \mathrm{CHCl}_{3}\right)$.

## 2-((S)-3-((R)-2-nitro-1-phenylethyl)thiochroman-4-ylidene)malononitrile 3c:

General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $28.9 \mathrm{mg}(0.080 \mathrm{mmol})$ with $80 \%$ yield; $90 \%$ ee was determined by HPLC on AS column, $35 \% 2$-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}, \mathrm{Uv} 254 \mathrm{~nm}, \mathrm{t}_{\mathrm{minor}}=$ $15.78 \mathrm{~min}, \mathrm{t}_{\text {major }}=31.04 \mathrm{~min}$.

## 2-((S)-2-((R)-2-nitro-1-phenylethyl)cyclohexylidene)malononitrile 3d:

General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $16.5 \mathrm{mg}(0.056 \mathrm{mmol})$ with $56 \%$ yield; $28 \%$ ee was determined by HPLC on AS column, $30 \% 2$-propanol $/$ hexane, $1.0 \mathrm{ml} / \mathrm{min}$, Uv $254 \mathrm{~nm}, \mathrm{t}_{\text {minor }}=7.3$ $\min , \mathrm{t}_{\text {major }}=9.3 \mathrm{~min}$.

## 2-((S)-2-((R)-1-(2-fluorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile 3aa:

General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $28.9 \mathrm{mg}(0.080 \mathrm{mmol})$ with $80 \%$ yield; $85 \%$ ee was determined by HPLC on AS column, $35 \% 2$-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}$, Uv $254 \mathrm{~nm}, \mathrm{t}_{\text {minor }}=$ $19.6 \mathrm{~min}, \mathrm{t}_{\text {major }}=23.3 \mathrm{~min}$.

## 2-((S)-2-((R)-1-(2-chlorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile 3ab:

General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid 29.4 mg ( 0.078 mmol ) with $78 \%$ yield; $90 \%$ ee was determined by HPLC on AS column, $35 \%$ 2-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}, \mathrm{Uv} 254 \mathrm{~nm}, \mathrm{t}_{\text {minor }}=8.1$ $\min , \mathrm{t}_{\text {major }}=10.1 \mathrm{~min}$.
2-((S)-2-((R)-1-(2-bromophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile 3ac:

General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $35 \mathrm{mg}(0.083 \mathrm{mmol})$ with $93 \%$ yield; $86 \%$ ee was determined by HPLC on AS column, $35 \% 2$-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}$, Uv $254 \mathrm{~nm}, \mathrm{t}_{\text {minor }}=$ $19.0 \mathrm{~min}, \mathrm{t}_{\text {major }}=19.9 \mathrm{~min}$.

## 2-((S)-2-((R)-1-(3-chlorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile 3ad:

General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $30.5 \mathrm{mg}(0.081 \mathrm{mmol})$ with $81 \%$ yield; $88 \%$ ee was determined by HPLC on AS column, $35 \%$ 2-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}$, $\mathrm{Uv} 254 \mathrm{~nm}, \mathrm{t}_{\text {minor }}=$ $10.3 \mathrm{~min}, \mathrm{t}_{\text {major }}=15.8 \mathrm{~min}$.

## 2-((S)-2-((R)-1-(3-methoxyphenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile 3ae:

General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $28.0 \mathrm{mg}(0.075 \mathrm{mmol})$ with $75 \%$ yield; $84 \%$ ee was determined by HPLC on AS column, $35 \%$ 2-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}$, Uv $254 \mathrm{~nm}, \mathrm{t}_{\text {minor }}=$ $29.5 \mathrm{~min}, \mathrm{t}_{\text {major }}=27.11 \mathrm{~min}$.

## 2-((S)-2-((R)-1-(4-chlorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-

 ylidene)malononitrile 3af:General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $27.1 \mathrm{mg}(0.072 \mathrm{mmol})$ with $72 \%$ yield; $88 \%$ ee was determined by HPLC on AS column, $30 \%$ 2-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}$, UV $254 \mathrm{~nm}, \mathrm{t}_{\text {minor }}=13.5$ $\mathrm{min}, \mathrm{t}_{\text {major }}=18.6 \mathrm{~min}$.
2-((S)-2-((R)-1-(4-bromophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile 3ag:

General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $30.4(0.072 \mathrm{mmol})$ with $72 \%$ yield; $92 \%$ ee was determined by HPLC on AS column, $30 \%$ 2-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}$, UV 254 nm , $\mathrm{t}_{\text {minor }}=$ $14.0 \mathrm{~min}, \mathrm{t}_{\text {major }}=19.4 \mathrm{~min}$.

## 2-((S)-2-((R)-1-(4-fluorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-

 ylidene)malononitrile 3ah:General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $29.2 \mathrm{mg}(0.081 \mathrm{mmol})$ with $81 \%$ yield; $87 \%$ ee was determined by HPLC on AS column, 30\% 2-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}$, UV $254 \mathrm{~nm}, \mathrm{t}_{\text {minor }}=16.8$ $\mathrm{min}, \mathrm{t}_{\text {major }}=25.4 \mathrm{~min}$.

## 2-((S)-3-((R)-1-(4-methoxyphenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile 3bh:

General experimental procedure I was followed to prepare the Michael addtion product 3a. The desired product was obtained as white solid $28.5 \mathrm{mg}(0.076 \mathrm{mmol})$ with $76 \%$ yield. $91 \%$ ee was determined by HPLC on AS column, $80 \%$ 2-propanol/hexane, $1.0 \mathrm{ml} / \mathrm{min}$, UV $254 \mathrm{~nm}, \mathrm{t}_{\text {minor }}=10.2$ $\mathrm{min}, \mathrm{t}_{\text {major }}=13.2 \mathrm{~min}$.

## 2-((S)-3-((R)-1-(2-fluorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile

General experimental procedure II was followed to prepare the Michael/hemiketalization
 product 3bb. The desired product was obtained as white solid with $75 \%$
 yield, mp: $183-185^{\circ} \mathrm{C}$; ${ }^{1} \mathrm{H}$ NMR $\left(500 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta(\mathrm{ppm}) 8.27(\mathrm{dd}, J$ $=1.5,8.0 \mathrm{~Hz}, 1 \mathrm{H}), 7.63(\mathrm{td}, J=1.5,6.0 \mathrm{~Hz}, 1 \mathrm{H}), 7.44-7.38(\mathrm{~m}, 1 \mathrm{H})$, 7.29-7.22 (m, 2H), 7.19-7.19 (m, 3H), $7.16(\mathrm{dd}, J=1.5,10 \mathrm{~Hz}, 1 \mathrm{H}), 5.02$ (t, $J=7.5 \mathrm{~Hz}, 1 \mathrm{H}$ ), 4.57 (dd, $J=10.5,18.5 \mathrm{~Hz}, 1 \mathrm{H}), 4.16$ (dd, $J=3.5$, $12.5 \mathrm{~Hz}, 2 \mathrm{H}), 3.58(\mathrm{~d}, J=8.5 \mathrm{~Hz}) ;{ }^{13} \mathrm{C}$ NMR $\left(125 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta(\mathrm{ppm})$
$165.0,155.9,137.6,131.1,127.9,125.3,122.4,118.4,116.8,116.6,114.8,112.8,79.4,76.3$,
66.8, 59.5; HRMS (ESI) calcd for $\mathrm{C}_{20} \mathrm{H}_{14} \mathrm{FN}_{3} \mathrm{O}_{3}[\mathrm{M}+\mathrm{H}]^{+}: 386.0911$, found: 386.0922; IR (KBr): v 3457, 2928, 2230, 1560, 1542, 1487, 1447, 1378, 1082, 1014, 832, $760 \mathrm{~cm}^{-1} ; 86 \%$ ee was determined by HPLC on ADH column, 10/90\% 2-propanol/hexane, $1.0 \mathrm{~mL} / \mathrm{min}$, UV 254 nm , $t_{\text {minor }}=19.1 \mathrm{~min}, t_{\text {major }}=21.8 \mathrm{~min}$.

## 2-((S)-3-((R)-1-(3-chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile:



General experimental procedure II was followed to prepare the Michael addition product 3bc. The desired product was obtained as white solid with $82 \%$ yield, $\mathrm{mp}: 187-189^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(500 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta=8.28$ (dd, $J=1.6,8.2 \mathrm{~Hz}, 1 \mathrm{H}$ ), 7.65 (ddd, $J=1.4,7.2,8.4 \mathrm{~Hz}, 1 \mathrm{H}$ ), $7.44-$ 7.38 (m, 2 H ), $7.36-7.31(\mathrm{~m}, 1 \mathrm{H}), 7.27-7.23(\mathrm{~m}, 1 \mathrm{H}), 7.18(\mathrm{dt}, J=$ $1.3,7.7 \mathrm{~Hz}, 1 \mathrm{H}), 7.10(\mathrm{dd}, J=1.1,8.4 \mathrm{~Hz}, 1 \mathrm{H}), 4.85(\mathrm{dd}, J=10.4,13.2 \mathrm{~Hz}, 1 \mathrm{H}), 4.52(\mathrm{dd}, J=$ $5.2,13.1 \mathrm{~Hz}, 1 \mathrm{H}), 4.18(\mathrm{dd}, J=2.4,12.5 \mathrm{~Hz}, 1 \mathrm{H}), 4.09(\mathrm{dd}, J=1.7,12.5 \mathrm{~Hz}, 1 \mathrm{H}), 3.73$ (dt, $J$ $=5.2,11.0 \mathrm{~Hz}, 1 \mathrm{H}), 3.32(\mathrm{td}, J=1.9,11.3 \mathrm{~Hz}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(126 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta=164.7$, $156.0,137.8,137.5,135.6,131.0,130.0,129.4,128.4,127.9,122.6,118.6,114.8,113.0,112.6$, 79.2, 77.1, 66.5, 43.2, 42.7. HRMS (ESI) calcd for $\mathrm{C}_{20} \mathrm{H}_{14} \mathrm{ClN}_{3} \mathrm{O}_{3}[\mathrm{M}+\mathrm{H}]^{+}: 380.0796$, found: 386.0809; IR (KBr): v 3470, 2225, 1572, 1556, 1497, 1363, 1094, 1010, 827, 784, 736; 88\% ee was determined by HPLC on ODH column, $10 / 90 \%$ 2-propanol/hexane, $1.0 \mathrm{~mL} / \mathrm{min}$, UV $254 \mathrm{~nm}, t_{\text {minor }}=18.3 \mathrm{~min}, t_{\text {major }}=20.8 \mathrm{~min}$.

## 2-((S)-3-((R)-1-(4-chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile:

General experimental procedure II was followed to prepare the Michael addtion product 3bd.
 The desired product was obtained as white solid with $80 \%$ yield, mp : $177-179{ }^{\circ} \mathrm{C}$; ${ }^{1} \mathrm{H}$ NMR $\left(500 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta=8.28(\mathrm{dd}, J=1.6,8.2 \mathrm{~Hz}$, 1 H ), 7.64 (ddd, $J=1.4,7.2,8.4 \mathrm{~Hz}, 1 \mathrm{H}), 7.46-7.41$ (m, 2 H ), 7.29 (d, $J=8.5 \mathrm{~Hz}, 2 \mathrm{H}$ ), $7.21-7.15$ (m, 1 H ), 7.09 (dd, $J=0.9,8.5 \mathrm{~Hz}, 1$ H), 4.84 (dd, $J=10.7,12.9 \mathrm{~Hz}, 1 \mathrm{H}$ ), $4.51(\mathrm{dd}, J=5.2,13.1 \mathrm{~Hz}, 1 \mathrm{H})$, 4.23-4.12 (m, 1 H), 4.12-4.03(m, 1H), $3.74(\mathrm{dt}, J=5.0,11.0 \mathrm{~Hz}, 1 \mathrm{H}), 3.29(\mathrm{td}, J=1.9,11.4$ $\mathrm{Hz}, 1 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( $125 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta=164.8,156.0,137.8,135.2,135.1,133.9,129.9$, 129.5, 127.9, 122.6, 118.6, 114.8, 113.1, 112.7, 79.1, 77.2, 66.5, 43.0. HRMS (ESI) calcd for $\mathrm{C}_{20} \mathrm{H}_{14} \mathrm{ClN}_{3} \mathrm{O}_{3}[\mathrm{M}+\mathrm{H}]^{+}: 380.0796$, found: 386.0802 . IR (KBr): v 3468, 2235, 1580, 1560, 1499, $1365,1097,1015,829,784,735 \mathrm{~cm}^{-1} ; 93 \%$ ee was determined by HPLC on AS column, $15 / 85 \%$ 2 -propanol/hexane, $1.0 \mathrm{~mL} / \mathrm{min}, \mathrm{UV} 254 \mathrm{~nm}, t_{\mathrm{minor}}=29.5 \mathrm{~min}, t_{\text {major }}=32.3 \mathrm{~min}$.

## 2-((S)-3-((R)-1-(3-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile:



General experimental procedure II was followed to prepare the Michael addition product 3be. The desired product was obtained as white solid with $79 \%$ yield, $\mathrm{mp}: 182-185{ }^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $\left(500 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta=8.28$ (dd, $J=1.6,8.2 \mathrm{~Hz}, 1 \mathrm{H}$ ), 7.64 (ddd, $J=1.4,7.2,8.4 \mathrm{~Hz}, 1 \mathrm{H}$ ), 7.55 (qd, $J=1.1,7.9 \mathrm{~Hz}, 1 \mathrm{H}), 7.48(\mathrm{t}, J=1.7 \mathrm{~Hz}, 1 \mathrm{H}), 7.37-7.32(\mathrm{~m}, 1$ H), $7.30(\mathrm{t}, J=1.4 \mathrm{~Hz}, 1 \mathrm{H}), 7.18(\mathrm{ddd}, J=0.9,7.3,8.2 \mathrm{~Hz}, 1 \mathrm{H}), 7.10(\mathrm{dd}, J=1.1,8.4 \mathrm{~Hz}, 1$ H), $4.84(\mathrm{dd}, J=10.6,13.1 \mathrm{~Hz}, 1 \mathrm{H}), 4.52(\mathrm{dd}, J=5.0,13.2 \mathrm{~Hz}, 1 \mathrm{H}), 4.18(\mathrm{dd}, J=2.4,12.5$ $\mathrm{Hz}, 1 \mathrm{H}), 4.09(\mathrm{dd}, J=1.7,12.5 \mathrm{~Hz}, 1 \mathrm{H}), 3.72(\mathrm{dt}, J=5.2,11.0 \mathrm{~Hz}, 1 \mathrm{H}), 3.32(\mathrm{td}, J=2.0,11.4$ $\mathrm{Hz}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $125 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta=164.7,156.0,137.8,137.8,132.4,131.3,131.2$, 127.9, 126.6, 123.7, 122.6, 118.6, 114.8, 113.0, 112.6, 79.2, 77.1, 66.5, 43.1, 42. HRMS (ESI)
calcd for $\mathrm{C}_{20} \mathrm{H}_{14} \mathrm{BrN}_{3} \mathrm{O}_{3}[\mathrm{M}+\mathrm{Na}]^{+}: 446.0110$, found: 446.0121; IR (KBr): 3468, 2928, 2232, $1585,1561,1494,1437,1375,1094,1015,830,768,738,641 ; 87 \%$ ee was determined by HPLC on ODH column, 20/80\% 2-propanol/hexane, $1.0 \mathrm{~mL} / \mathrm{min}$, UV $254 \mathrm{~nm}, t_{\text {minor }}=17.6 \mathrm{~min}$, $t_{\text {major }}=21.9 \mathrm{~min}$.

## 2-((S)-3-((R)-1-(4-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile



General experimental procedure II was followed to prepare the Michael addition product $\mathbf{3 b f}$. The desired product was obtained as white solid with $83 \%$ yield, $\mathrm{mp}: 178-180{ }^{\circ} \mathrm{C}$; ${ }^{1} \mathrm{H}$ NMR ( $500 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta=8.28$ (dd, $J=1.3,8.2 \mathrm{~Hz}, 1 \mathrm{H}$ ), 7.64 (ddd, $J=1.6,7.1,8.4 \mathrm{~Hz}, 1 \mathrm{H}), 7.62-$ 7.56 (m, 2 H), 7.26-7.20 (m, 2 H$), 7.20-7.15(\mathrm{~m}, 1 \mathrm{H}), 7.09$ (dd, $J=$ $0.9,8.5 \mathrm{~Hz}, 1 \mathrm{H}), 4.84(\mathrm{dd}, J=10.7,12.9 \mathrm{~Hz}, 1 \mathrm{H}), 4.51(\mathrm{dd}, J=5.0,12.9 \mathrm{~Hz}, 1 \mathrm{H}), 4.16(\mathrm{dd}, J$ $=2.4,12.5 \mathrm{~Hz}, 1 \mathrm{H}), 4.12-4.05(\mathrm{~m}, 1 \mathrm{H}), 3.73(\mathrm{dt}, J=5.0,11.0 \mathrm{~Hz}, 1 \mathrm{H}), 3.29(\mathrm{td}, J=2.0,11.4$ $\mathrm{Hz}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $125 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta=164.8,156.0,137.8,134.4,132.9,129.7$, 127.9, 123.3, 122.6, 118.6, 114.8, 113.1, 112.7, 79.1, 77.2, 66.5, 43.0, 42.7. HRMS (ESI) calcd for $\mathrm{C}_{20} \mathrm{H}_{14} \mathrm{BrN}_{3} \mathrm{O}_{3}[\mathrm{M}+\mathrm{Na}]^{+}: 446.0110$, found: 446.0121; IR (KBr): v 3466, 2929, 2234, 1587, 1564, 1497, 1439, 1379, 1095, 1017, 832, 770, 740, $641 \mathrm{~cm}^{-1} ; 86 \%$ ee was determined by HPLC on ODH column, $10 / 90 \%$ 2-propanol $/$ hexane, $1.0 \mathrm{~mL} / \mathrm{min}$, UV $254 \mathrm{~nm}, t_{\text {minor }}=12.6 \mathrm{~min}, t_{\text {major }}=18.6$ min.

2-(2-(1-(4-Methylphenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)-ylidene)malononitrile. General experimental procedure II was followed to prepare the
 Michael addition product 3bg. The desired product was obtained as white solid with $81 \%$ yield mp : $183-185^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR $(500 \mathrm{MHz}$, $\mathrm{CDCl}_{3}$ ) $\delta(\mathrm{ppm}) 8.26(\mathrm{dd}, J=1.0,8.0 \mathrm{~Hz}, 1 \mathrm{H}), 7.63(\mathrm{td}, J=7.5$, $0.5 \mathrm{~Hz}, 1 \mathrm{H}), 7.29-7.26(\mathrm{~m}, 4 \mathrm{H}), 7.17(\mathrm{td}, J=1,7.0 \mathrm{~Hz}, 1 \mathrm{H}), 7.07$ (dd, $J=1.0,8.5 \mathrm{~Hz}, 1 \mathrm{H}), 4.84(\mathrm{dd}, J=2.5,13.0 \mathrm{~Hz}, 1 \mathrm{H}), 4.49(\mathrm{dd}$, $J=5.5,12.5 \mathrm{~Hz}, 1 \mathrm{H}), 4.12-4.11(\mathrm{~m}, 2 \mathrm{H}), 3.69(\mathrm{dt}, J=3,11.5 \mathrm{~Hz}$, $1 \mathrm{H}), 3.30\left(\mathrm{dd}, J=2,13.5 \mathrm{~Hz}, 1 \mathrm{H} ;{ }^{13} \mathrm{C}\right.$ NMR ( $125 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta(\mathrm{ppm}) 174.4,166.0,156.1$, 139. 9, 137.7, 132.2, 131.7, 130.3, 127.9, 122.4, 114.9, 78.9, 77.3, 66.6, 43.3, 43.1, 21.1; HRMS (ESI) calcd for $\mathrm{C}_{21} \mathrm{H}_{17} \mathrm{~N}_{3} \mathrm{O}_{3}[\mathrm{M}+\mathrm{H}]^{+}: 360.1342$, found: 360.1331; IR (KBr): 3468, 2230, 1574, $1559,1515,1378,1260,1180,1037,828,775,751 \mathrm{~cm}-1 ; 90 \%$ ee was determined by HPLC on ADH column, $15 / 85 \%$ 2-propanol $/$ hexane, $1.0 \mathrm{~mL} / \mathrm{min}$, UV $254 \mathrm{~nm}, t_{\text {minor }}=21.5 \mathrm{~min}, t_{\text {major }}=26.4$ min.

## (R)-3-((R)-2-nitro-1-phenylethyl)chroman-4-one:



To a stirred solution of 3ba (1 equiv) in acetone and water as solvent, $\mathrm{KMnO}_{4}$ (2 equiv) was added. The solution was stirred at ambient temperature for 4 h . After the reaction was completed (monitored by TLC), the resulting mixture was concentrated under reduced pressure and the residue was purified through column chromatography on silica gel to give the product $\mathbf{7 b}$. The desired product 7b was obtained as white solid with $67 \%$ yield; mp : $155-$ $157^{\circ} \mathrm{C} ;{ }^{1} \mathrm{H}$ NMR ( $500 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta=7.95(\mathrm{dd}, J=1.9,7.9 \mathrm{~Hz}, 1 \mathrm{H}), 7.56$ (ddd, $J=1.7,7.1$, $8.5 \mathrm{~Hz}, 1 \mathrm{H}), 7.44-7.40(\mathrm{~m}, 2 \mathrm{H}), 7.38-7.34(\mathrm{~m}, 1 \mathrm{H}), 7.33-7.30(\mathrm{~m}, 2 \mathrm{H}), 7.13-7.08(\mathrm{~m}, 1$ H), $7.04-7.00(\mathrm{~m}, 1 \mathrm{H}), 4.96(\mathrm{dd}, J=5.0,13.2 \mathrm{~Hz}, 1 \mathrm{H}), 4.82(\mathrm{dd}, J=10.6,13.4 \mathrm{~Hz}, 1 \mathrm{H}), 4.28$
$(\mathrm{dd}, J=3.5,11.7 \mathrm{~Hz}, 1 \mathrm{H}), 4.04(\mathrm{dd}, J=5.2,11.8 \mathrm{~Hz}, 1 \mathrm{H}), 3.95(\mathrm{dt}, J=4.7,10.9 \mathrm{~Hz}, 1 \mathrm{H})$, 2.84-2.78 (m, 1 H); ${ }^{13} \mathrm{C}$ NMR (125 MHz, $\left.\mathrm{CDCl}_{3}\right) \delta=192.8,161.4,136.7,136.3,129.4,128.5$, $128.0,127.7,122.0,120.0,117.9,78.5,68.7,48.6,41.2$; HRMS (ESI) calcd for $\mathrm{C}_{17} \mathrm{H}_{15} \mathrm{NO}_{4}$ $[\mathrm{M}+\mathrm{H}]^{+}: 298.1074$, found: 298.1087; IR (KBr): 3467, 2228, 1779, 1575, 1560, 1525, 1381, $1265,1182,1034,820,770 \mathrm{~cm}^{-1}$;
(3R,3aS,9bR)-3-phenyl-2,3,3a,4,5,9b-hexahydro-1H-benzo[g]indole:
To a stirred solution of $7 \mathbf{7 a}$ ( 1 equiv) in acetic acid as solvent, Zn dust ( 10 equiv) was added. The
 solution was stirred at ambient temperature for 8 h . After the reaction was completed (monitored by TLC), the resulting mixture was concentrated under reduced pressure and the residue was purified through column chromatography on silica gel to give the product $\mathbf{8 a}$. The desired product was obtained as pasty solid with $61 \%$ yield; ${ }^{1} \mathrm{H} \operatorname{NMR}\left(500 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta=7.47(\mathrm{~d}, J=7.3 \mathrm{~Hz}$, $1 \mathrm{H}), 7.37-7.29(\mathrm{~m}, 4 \mathrm{H}), 7.25-7.10(\mathrm{~m}, 4 \mathrm{H}), 4.45(\mathrm{~d}, J=8.2 \mathrm{~Hz}, 1 \mathrm{H}), 3.49(\mathrm{dd}, J=6.8,10.6$ Hz, 1 H), 3.09-3.04 (m, 1 H), 3.04-2.98 (m, 1 H), 2.84-2.77 (m, 1 H ), 2.64 (ddd, $J=4.1,9.1$, $15.8 \mathrm{~Hz}, 1 \mathrm{H}), 2.55-2.45(\mathrm{~m}, 1 \mathrm{H}), 1.95-1.88(\mathrm{~m}, 1 \mathrm{H}), 1.60(\mathrm{dtd}, J=4.1,8.8,13.2 \mathrm{~Hz}, 1 \mathrm{H})$; ${ }^{13} \mathrm{C}$ NMR $\left(125 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta=144.5,138.4,138.2,129.7,128.9,128.4,127.9,126.7,126.5$, $126.4,59.7,55.1,52.6,46.9,28.3,26.7$; HRMS (ESI) calcd for $\mathrm{C}_{18} \mathrm{H}_{19} \mathrm{~N}[\mathrm{M}+\mathrm{Na}]^{+}: 298.1074$, found: 298.1087; IR (KBr): 3625, 3650, 2230, 1625, 1556, 1378, 1271, 1185, 1037, 838, 779 $\mathrm{cm}^{-1} ; 85 \%$ ee was determined by HPLC on phenomenex Amylose 2 column, 20/80\% 2propanol $/$ hexane, $1.0 \mathrm{~mL} / \mathrm{min}$, UV $254 \mathrm{~nm}, t_{\text {minor }}=19.6 \mathrm{~min}, t_{\text {major }}=22.3 \mathrm{~min}$.

## (3R,3aS,9bR)-3-phenyl-1,2,3,3a,4,9b-hexahydrochromeno[4,3-b]pyrrole:



To a stirred solution of $\mathbf{7 b}$ ( 1 equiv) in acetic acid as solvent, Zn dust (10 equiv) was added. The solution was stirred at ambient temperature for 8 h . After the reaction was completed (monitored by TLC), the resulting mixture was concentrated under reduced pressure and the residue was purified through column chromatography on silica gel to give the product $\mathbf{8 b}$. The desired product $\mathbf{8 b}$ was obtained as pasty solid with $64 \%$ yield; ${ }^{1} \mathrm{H}$ NMR (500MHz, DMSO-d $\left.{ }_{6}\right) \delta=7.40(\mathrm{dd}, J=1.6,7.6 \mathrm{~Hz}, 1 \mathrm{H}), 7.36-7.30(\mathrm{~m}, 4 \mathrm{H}), 7.25-7.20$ $(\mathrm{m}, 1 \mathrm{H}), 7.18-7.12(\mathrm{~m}, 1 \mathrm{H}), 6.94(\mathrm{dt}, J=1.3,7.4 \mathrm{~Hz}, 1 \mathrm{H}), 6.84(\mathrm{dd}, J=1.3,8.2 \mathrm{~Hz}, 1 \mathrm{H})$, $4.23-4.12(\mathrm{~m}, 2 \mathrm{H}), 3.67(\mathrm{dd}, J=9.8,10.7 \mathrm{~Hz}, 1 \mathrm{H}), 3.31(\mathrm{dd}, J=7.9,10.7 \mathrm{~Hz}, 1 \mathrm{H}), 2.88(\mathrm{~d}, J$ $=5.4 \mathrm{~Hz}, 1 \mathrm{H}), 2.77(\mathrm{dd}, J=8.7,10.6 \mathrm{~Hz}, 1 \mathrm{H}), 2.43(\mathrm{tdd}, J=5.1,6.9,9.8 \mathrm{~Hz}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR (125 MHz, DMSO- $\mathrm{d}_{6}$ ) $\delta=155.1,144.4,131.1,129.0,128.3,128.0,126.7,125.1,121.2,116.9$, $66.0,56.3,55.3,48.3,45.8$; HRMS (ESI) calcd for $\mathrm{C}_{17} \mathrm{H}_{17} \mathrm{NO}[\mathrm{M}+\mathrm{H}]^{+}: 252.1383$, found: 252.1370; IR (KBr): 3628, 3655, 2235, 1628, 1558, 1374, 1275, 1182, 1039, 845, $740 \mathrm{~cm}^{-1}$; $75 \%$ ee was determined by HPLC on ODH column, $30 / 70 \%$ 2-propanol/hexane, $1.0 \mathrm{~mL} / \mathrm{min}$, UV $254 \mathrm{~nm}, t_{\text {minor }}=49.6 \mathrm{~min}, t_{\text {major }}=59.0 \mathrm{~min}$.

References:

1) D. Xue, Y.-C. Chen, Q.-W. Wang, L.-F. Cun, J. Zhu, J.-G. Deng, Org. Lett. 2005, 7, 5293-5296;
2) T. B. Poulsen, M. Bell, K. A. Jorgensen, Org. Biomol. Chem. 2006, 4, 63-70.
3) Zhou, L.-H.; Wang, N.; Chen, G.-N.; Yang, Q.; Yang, S.-Y.; Zhang, W.; Zhang, Y.; Yu, X.-Q. Journal of Molecular Catalysis B: Enzymatic 2014, 109, 170-177
${ }^{1}$ H NMR of 2-(2-(1-(2-fluorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile

${ }^{13}$ C NMR of 2-(2-(1-(4-fluorophenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile

${ }^{1}$ H NMR of 2-(3-(1-(3-Chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.



${ }^{13}$ C NMR of 2-(3-(1-(3-Chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.

${ }^{1}$ H NMR of 2-(3-(1-(4-chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.



${ }^{13}$ C NMR of 2-(3-(1-(4-chlorophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.

DM-19-133L[9CI]. ....... Vishwanath,



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|  |



${ }^{1}$ H NMR of 2-(3-(1-(3-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.



${ }^{13}$ C NMR of 2-(3-(1-(3-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.

${ }^{1} \mathrm{H}$ NMR of 2-(3-(1-(4-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.



${ }^{13}$ C NMR of 2-(3-(1-(4-bromophenyl)-2-nitroethyl)chroman-4-ylidene)malononitrile.
$\qquad$ Vishwanath,



${ }^{1}$ H NMR of 2-(2-(1-(4-Methylphenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile


${ }^{13}$ C NMR of 2-(2-(1-(4-Methylphenyl)-2-nitroethyl)-3,4-dihydronaphthalen-1(2H)ylidene)malononitrile

${ }^{1} H$ NMR of (R)-3-((R)-2-nitro-1-phenylethyl)chroman-4-one.

## DM-25-159........... Vishwanatl



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ô
${ }^{13}$ C NMR of (R)-3-((R)-2-nitro-1-phenylethyl)chroman-4-one

${ }^{1}$ H NMR of (3R,3aS,9bR)-3-phenyl-2,3,3a,4,5,9b-hexahydro-1H-benzo[g]indole.

DM-25-151.........Vishwanath,

${ }^{13}$ C NMR of (3R,3aS,9bR)-3-phenyl-2,3,3a,4,5,9b-hexahydro-1H-benzo[g]indole.


## NOESY spectrum of (3R,3aS,9bR)-3-phenyl-2,3,3a,4,5,9b-hexahydro-1H-benzo[g]indole.



NOESY spectum shows no spatial interactions between Ha and Hc , which indicates those protons are in "anti" configuration in cycl ised product.
${ }^{1}$ H NMR of (3R.3aS.9bR)-3-phenvl-1.2.3.3a.4.9b-hexahvdrochromenol4.3-blbvrrole DM-25-161...........Vishwanath



${ }^{13}$ C NMR of (3R,3aS,9bR)-3-phenyl-1,2,3,3a,4,9b-hexahydrochromeno[4,3-b]pyrrole.







## HPLC profile for table3, entry 1


PDA Ch1 254 nm 4 nm

| PeakTable |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 1 | 13.955 | 26947195 | 735911 | 49.599 | 70.545 |
| 2 | 25.135 | 27383159 | 307271 | 50.401 | 29.455 |
| Total |  | 54330354 | 1043181 | 100.000 | 100.000 |



A Multi $1 / 254 n m 4 n m$
PDA Ch1 254 nm 4 nm

| Peak\# | Ret. TimeakTable |  |  |  |  |
| ---: | :---: | :---: | ---: | ---: | ---: |
| 1 | 13.675 | Area | Height | Area $\%$ | Height $\%$ |
| 2 | 2569671 | 75341 | 4.419 | 10.609 |  |
| Total. |  | 55575036 | 634836 | 95.581 | 89.391 |

## HPLC profile for table3, entry 2


)A Multi $1 / 220 \mathrm{~nm} 4 \mathrm{~nm}$
PDA Ch1 220 nm 4 nm

| Peak | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 15.081 | 9015261 | 151860 | 49.881 | 46.087 |
| 2 | 17.564 | 9058285 | 177646 | 50.119 | 53.913 |
| Total |  | 18073545 | 329505 | 100.000 | 100.000 |



A Multi $1 / 220 \mathrm{~nm} 4 \mathrm{~nm}$
PDA. Ch1 220 nm 4 nm

| PeakTable |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak $\#$ | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 1 | 15.141 | 2168839 | 36067 | 2.366 | 1.938 |
| 2 | 17.587 | 89482351 | 1824522 | 97.634 | 98.062 |
| Totai. |  | 91651190 | 1860589 | 100.000 | 100.000 |

## HPLC profile for table3, entry 3



3 t .A Ch1/310nm
Petector A Ch1 310nm
De|r|r|r|

| Peak\# | Ret. Time | Area | Height | Area \% |
| ---: | ---: | ---: | ---: | ---: |
| 1 | 11.574 | 1261592 | 49068 | 50.309 |
| 2 | 20.659 | 1246085 | 22342 | 49.691 |
| Total |  | 2507677 | 71411 | 100.000 |


| Height \% |
| ---: |
| 68.713 |
| 31.287 |
| 100.000 |


Detector A Ch1 310nm

| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 11.502 | 3806054 | 149489 | 4.875 | 11.178 |
| 2 | 19.914 | 74274536 | 1187841 | 95.125 | 88.822 |
| Total |  | 78080590 | 1337330 | 100.000 | 100.000 |

HPLC profile for table3, entry 4



## HPLC profile for table 4, entry 1




## HPLC profile for table 4, entry 2



Jet.A Ch1/254nm
Detector A Ch1 254 nm

|  | PeakTable |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height \% |
| 1 | 8.258 | 11288799 | 446150 | 49.533 | 53.127 |
| 2 | 10.260 | 11501444 | 393635 | 50.467 | 46.873 |
| Total |  | 22790243 | 839784 | 100.000 | 100.000 |


et.A Ch1/254nm
Detector A Ch1 254nm

| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 8.169 | 464727 | 21637 | 5.208 | 6.442 |
| 2 | 10.168 | 8458352 | 314233 | 94.792 | 93.558 |
| 3 | 14.933 | 0 | 0 | 0.000 | 0.000 |
| Total |  | 8923080 | 335871 | 100.000 | 100.000 |

## HPLC profile for table 4, entry 3



| Detector A Chi 254nm |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Peak\# | Ret. Time | Area | Height | Area \% | Height \% |
| 1 | 19.536 | 294078 | 10079 | 49.235 | 51.411 |
| 2 | 20.468 | 303213 | 9526 | 50.765 | 48.589 |
| Total |  | 597291 | 19604 | 100.000 | 100.000 |


PeakTable

| Detector A.Ch1 254nm |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| $\|c\|$ Peak\# Ret. Time Area Height | Area \% | Height \% |  |  |  |
| 1 | 19.039 | 83939 | 3156 | 3.965 | 4.696 |
| 2 | 19.934 | 2033259 | 64049 | 96.035 | 95.304 |
| Total |  | 2117199 | 67205 | 100.000 | 100.000 |

HPLC profile for table 4, entry 4

t.A Ch1/254nm

## PeakTable

Detector A Ch1 254 nm

| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 10.241 | 7678770 | 338919 | 49.292 | 68.827 |
| 2 | 15.809 | 7899399 | 153502 | 50.708 | 31.173 |
| Totai. |  | 15578168 | 492421 | 100.000 | 100.000 |


Detector A Ch1 254 nm

| PeakTable |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak\# | Ret. Time | Area | Height | Area \% | Height \% |
| 1 | 10.357 | 1097625 | 55457 | 5.789 | 12.821 |
| 2 | 15.816 | 17861565 | 377079 | 94.211 | 87.179 |
| Totail |  | 18959190 | 432537 | 100.000 | 100.000 |

## HPLC profile for table 4, entry 5



et.A Ch1/254nm

PeakTable
Detector A. Ch1 254 nm

| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 27.118 | 5895994 | 91631 | 91.538 | 91.357 |
| 2 | 29.587 | 545072 | 8669 | 8.462 | 8.643 |
| Total |  | 6441065 | 100300 | 100.000 | 100.000 |

HPLC profile for table 4, entry 6

let.A Ch $1 / 254 \mathrm{~nm}$

| PeakTable |  |  |  |  |  |
| :--- | :---: | :---: | :---: | :---: | :---: |
| Detector A Ch1 254nm      <br> Peak\# Ret. Time Area Height Area $\%$ Height $\%$ <br> 1 13.115 5624880 212978 50.629 65.124 <br> 2 18.162 5485166 114057 49.371 34.876 <br> Total  11110045 327034 100.000 100.000 |  |  |  |  |  |


Detector A Ch1 254nm

| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height \% |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 13.497 | 769089 | 28600 | 5.796 | 11.091 |
| 2 | 18.569 | 12499451 | 229261 | 94.204 | 88.909 |
| Totai |  | 13268540 | 257861 | 100.000 | 100.000 |

## HPLC profile for table 4, entry 7



2t.A Ch1/254nm

PeakTable
Detector A Ch1 254 nm

| PeakTable |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 1 | 14.157 | 2371480 | 82770 | 50.399 | 64.002 |
| 2 | 19.751 | 2333973 | 46554 | 49.601 | 35.998 |
| Total |  | 4705453 | 129324 | 100.000 | 100.000 |


at.A Ch1/254nm
Detector A Ch1 254nm

| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 14.054 | 781132 | 28397 | 4.364 | 8.213 |
| 2 | 19.384 | 17117417 | 317366 | 95.636 | 91.787 |
| Total. |  | 17898550 | 345763 | 100.000 | 100.000 |

HPLC profile for table 4, entry 8

Detector A Ch1 254nm

| PeakTable |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 1 | 14.157 | 2371480 | 82770 | 50.399 | 64.002 |
| 2 | 19.751 | 2333973 | 46554 | 49.601 | 35.998 |
| Total |  | 4705453 | 129324 | 100.000 | 100.000 |


st.A Ch1/254nm

|  |  | PeakTable |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Detector A Ch1 254 nm ( PeakTable |  |  |  |  |  |
| Peak\# | Ret. Time | Area | Height | Area \% | Height \% |
| 1 | 14.054 | 781132 | 28397 | 4.364 | 8.213 |
| 2 | 19.384 | 17117417 | 317366 | 95.636 | 91.787 |
| Total |  | 17898550 | 345763 | 100.000 | 100.000 |

HPLC profile for table 5, entry 1


JA Multi $1 / 220 \mathrm{~nm} 4 \mathrm{~nm}$
PDA Ch1 220 nm 4 nm

|  |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak $\#$ | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 1 | 15.081 | 9015261 | 151860 | 49.881 | 46.087 |
| 2 | 17.564 | 9058285 | 177646 | 50.119 | 53.913 |
| Total |  | 18073545 | 329505 | 100.000 | 100.000 |



A Multi $1 / 220 \mathrm{~nm} 4 \mathrm{~nm}$
PDA. Ch1 220 nm 4nm

| PeakTable |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 2 | 15.141 | 2168839 | 36067 | 2.366 | 1.938 |
| Totai | 17.587 | 89482351 | 1824522 | 97.634 | 98.062 |

HPLC profile for table 5, entry 2


A Multi $1 / 254 \mathrm{~nm} 4 \mathrm{~nm}$
PDA Ch1 254 nm 4nm

| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 18.361 | 3382550 | 100386 | 50.075 | 52.415 |
| 2 | 20.971 | 3372374 | 91134 | 49.925 | 47.585 |
| Total |  | 6754924 | 191519 | 100.000 | 100.000 |


PDA Ch1 254 nm 4nm

|  |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 1 | 19.103 | 662258 | 15482 | 6.821 | 6.384 |
| 2 | 21.807 | 9046242 | 227034 | 93.179 | 93.616 |
| Total |  | 9708500 | 242516 | 100.000 | 100.000 |

## HPLC profile for table 5, entry 3




IA Multi $1 / 220 \mathrm{~nm} 4 \mathrm{~nm}$
PeakTable
PDA. Ch1 220 nm 4 nm

| Peak $=~$ | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 18.368 | 3519944 | 69919 | 5.814 | 8.201 |
| 2 | 20.881 | 57027713 | 782674 | 94.186 | 91.799 |
| Tota. |  | 60547657 | 852593 | 100.000 | 100.000 |

HPLC profile for table 5, entry 4

Detector A Ch1 254nm

| PeakTable |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak\# | Ret. Time | Area | Height | Area \% | Height \% |
| 1 | 29.044 | 1893763 | 41025 | 50.683 | 51.835 |
| 2 | 31.797 | 1842691 | 38121 | 49.317 | 48.165 |
| Total. |  | 3736454 | 79146 | 100.000 | 100.000 |


Detector A Ch1 254 nm

| PeakTable |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak\# | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 1 | 29.566 | 475140 | 10639 | 3.767 | 4.294 |
| 2 | 32.348 | 12138613 | 237148 | 96.233 | 95.706 |
| Total. |  | 12613754 | 247787 | 100.000 | 100.000 |

HPLC profile for table 5, entry 5

Detector A Ch1 254 nm

|  |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak\# PeakTable |  |  |  |  |  |
| 1 | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 2 | 17.880 | 41183161 | 1334860 | 49.888 | 54.610 |
| Total | 22.301 | 41368840 | 1109496 | 50.112 | 45.390 |



## HPLC profile for table 5, entry 6


|A Multi $1 / 254 \mathrm{~nm} 4 \mathrm{~nm}$

## PeakTable

PDA Chl 254 mm 4 mm

| Peak | Ret. Time | Area | Height | Area \% | Height \% |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 12.523 | 2154083 | 73118 | 50.258 | 69.154 |
| 2 | 18.718 | 2131933 | 32614 | 49.742 | 30.846 |
| Total |  | 4286016 | 105732 | 100.000 | 100.000 |



DA Multi $1 / 254 \mathrm{~nm} 4 \mathrm{~nm}$
PDA. Ch1 254 nm 4nm

| Peak | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 | 12.615 | 774877 | 25984 | 7.263 | 15.334 |
| 2 | 18.640 | 9894663 | 143472 | 92.737 | 84.666 |
| Totai |  | 10669539 | 169455 | 100.000 | 100.000 |

## HPLC profile for table 5, entry 7



DA Multi $1 / 254 \mathrm{~nm} 4 \mathrm{~nm}$
PDA Ch1 254 nm 4nm

| Peak | Ret. Time | Area | Height | Area $\%$ |
| ---: | :---: | :---: | :---: | ---: |
| 1 | 21.743 | 1146070 | 20286 | 49.465 |
| 2 | 26.713 | 1170878 | 17472 | 50.535 |
| Total |  | 2316948 | 37758 | 100.000 |


| Height $\%$ |
| ---: |
| 53.727 |
| 46.273 |
| 100.000 |



DA Multi $1 / 254 \mathrm{~nm} 4 \mathrm{~nm}$
PDA Ch1 254 nm 4 nm

| PeakTable |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 1 | 21.587 | 190903 | 3889 | 5.209 | 7.228 |
| 2 | 26.497 | 3473651 | 49914 | 94.791 | 92.772 |
| Total |  | 3664554 | 53803 | 100.000 | 100.000 |

## HPLC profile for table 5, entry 8




JA Multi $1 / 254 \mathrm{~nm} 4 \mathrm{~nm}$
PDA. Ch1 254 nm 4 nm

| PeakTable |  |  |  |  |  |
| ---: | ---: | ---: | ---: | ---: | ---: |
| Peak | Ret. Time | Area | Height | Area $\%$ | Height $\%$ |
| 1 | 10.558 | 1416199 | 39060 | 9.831 | 10.357 |
| 2 | 13.269 | 12988798 | 338071 | 90.169 | 89.643 |
| Totai |  | 14404997 | 377131 | 100.000 | 100.000 |

## HPLC profile for Scheme 2, entry 8a



1 PDA Multi $1 / 254 \mathrm{~nm} 4 \mathrm{~nm}$

| PDA Chil 254 nm 4 nm PeakTable |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |
| Peak | Ret. Time | Area | Height | Area \% | Height \% |
| 1 | 18.361 | 3382550 | 100386 | 50.075 | 52.415 |
| 2 | 20.971 | 3372374 | 91134 | 49.925 | 47.585 |
| Total |  | 6754924 | 191519 | 100.000 | 100.000 |



1 PDA Multi $1 / 254 \mathrm{~nm} 4 \mathrm{~nm}$
PDACh1 254nm 4nm

| Peakik | Ret. Time | Area | Heaght | Area \% | Height \%a |
| ---: | :---: | ---: | ---: | ---: | ---: |
| 1 | 19.636 | 289704 | 7079 | 7.480 | 8.968 |
| 2 | 22.375 | 3583401 | 71856 | 92.520 | 91.032 |
| Total |  | 3873105 | 78936 | 100.000 | 100.000 |

## HPLC profile for Scheme 2, entry 8b


mAU


1. PDA Multi $3 / 254 \mathrm{~nm} 4 \mathrm{~nm}$

|  |  |  | PeakTat |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| PDACr3 254 mm 4 mm |  |  |  |  |  |
| Pesker | Ret Time | Asea | Hegir | Asea\% | Hagro |
| 1 | 40.651 | 20982936 | 230042 | 87.569 | 88691 |
| 2 | \$8.058 | 2978791 | 29334 | 12.431 | 11.309 |
| Toud |  | 2861727 | 259376 | 10000 | 10000 |

