



Targeting IGF1R pathway in cancer with microRNAs: How close are we?

Arathy S. Kumar, Suresh K. Rayala & Ganesh Venkatraman

To cite this article: Arathy S. Kumar, Suresh K. Rayala & Ganesh Venkatraman (2018) Targeting IGF1R pathway in cancer with microRNAs: How close are we?, RNA Biology, 15:3, 320-326, DOI: [10.1080/15476286.2017.1338240](https://doi.org/10.1080/15476286.2017.1338240)

To link to this article: <https://doi.org/10.1080/15476286.2017.1338240>



Published online: 01 Feb 2018.



Submit your article to this journal [↗](#)



Article views: 705



View related articles [↗](#)



View Crossmark data [↗](#)



Citing articles: 6 View citing articles [↗](#)

REVIEW



Targeting IGF1R pathway in cancer with microRNAs: How close are we?

Arathy S. Kumar^a, Suresh K. Rayala^a, and Ganesh Venkatraman^b

^aDepartment of Biotechnology, Indian Institute of Technology, Madras (IIT M), Chennai, India; ^bDepartment of Human Genetics, College of Biomedical Sciences, Technology & Research, Sri Ramachandra University, Porur, Chennai, India

ABSTRACT

Cancer of the head and neck are the most common cancers in India and account for 30% of all cancers. At molecular level, it could be attributed to the overexpression of growth factors like IGF1-R, EGFR, VEGF-R and deregulation of cell cycle regulators and tumor suppressors. IGF1-R is an emerging target in head and neck cancer treatment, because of its reported role in tumor development, progression and metastasis. IGF1R targeted agents are in advanced stages of clinical development. Nevertheless, these agents suffer from several disadvantages including acquired resistance and toxic side effects. Hence there is a need for developing newer agents targeting not only the receptor but also its downstream signaling. miRNAs are considered as master regulators of gene expression of multiple genes and has been widely reported to be a promising therapeutic strategy. This review discusses the present status of research in both these arenas and emphasizes the role of miRNA as a promising agent for biologic therapy.

ARTICLE HISTORY

Received 6 March 2017
Accepted 31 May 2017

KEYWORDS

Growth factor receptor signaling; head neck cancer; IGF1R; miR493; miRNA therapy

Introduction

Role of IGF1R signaling in multiple functions including proliferation, differentiation, glucose homeostasis, energy metabolism and its conserved regulatory module is an indication of its importance in coordinating growth, development and metabolism. IGF1 pathway comprise of its ligand- IGF1, respective receptor- IGF1R, ligand binding proteins- IGF1R, downstream adaptor molecules such Shc and insulin receptor substrate (IRS) –1, –2, –3, and –4 and downstream effectors- AKT1, MAPK, etc. Insulin-like growth factor 1 (IGF-1) is a peptide hormone, functions as the major mediator of growth hormone (GH)-stimulated somatic growth and GH-independent anabolic reactions in most cells and tissues. Circulation of IGF1 in serum is in its bound form to high affinity binding proteins [1]. Synthesis of IGF1 is in 2 different ways- synthesis in the liver and secreted into the blood which is under the control of growth hormones. IGF1 synthesis from peripheral tissues such as bone in autocrine/paracrine manner, which is under the control of growth hormones as well as by factors that are secreted locally by the surrounding cell types. These IGF1 also enter the systemic circulation, which is not under the regulation of GH and hence the regulated secretion of autocrine or paracrine secreted IGF1 and the factors responsible for the secretion needed to be studied and explored more [2].

IGF1R pathway is studied to have a role to play in prenatal and post natal growth in mice and in life span regulation. At the tissue level IGF1R mediated PI3K-AKT and/or ERKs mediates differentiation and also shown to maintain the myocardium and brain. Its role in regulation of vascular homeostasis

and endothelial function and reports showed decreased expression of IGF1R increases the fibrosis of kidney disease in mice models [3]. But there are paradoxical results showing IGF1R and longevity in lower animal forms such as *C.elegans* [4]. IGF1R interaction and cross talk with other growth factor receptors are studied to have a major role in cells; interactions between IR and IGF1R in muscle were found to have significance in glucose homeostasis in mice models [5].

IGF-1 uses its effects via binding followed by the activation of the IGF-1 receptor, which is a tyrosine kinase protein, which is widely distributed and it facilitates circulating IGF-1 to perform the function of balanced growth among multiple tissues and organs. IGF1R consists of 2 half-receptors with an extracellular α -subunit and one transmembrane β -subunit which perform tyrosine kinase activity on binding to the ligand. Receptor has affinity toward the other ligand IGF2 and the ability of IGF1R to form Insulin receptor (IR) and perform the signaling by binding to IGF1/2 makes the signaling complex. Binding of ligand to IGF1R results in the autophosphorylation of IGF1R at tyrosines 1131,1135, and 1136 in the kinase domain of β -subunit of the receptor. This phosphorylation induces further phosphorylation of juxtamembrane tyrosines and carboxyl-terminal serines creating docking sites for proteins including IR substrates 1 to 4 (IRS-1 to IRS-4), and Shc (Src homology and collagen domain protein). Binding of these molecules activates signaling via the phosphatidylinositol-3-kinase (PI3K)-AKT and RAS/RAF/mitogen-activated protein kinase (MAPK) pathways [6].

IGF1R in cancer: How and why?

Reports of deregulated expression of IGF1R pathway molecules including the receptor and downstream effectors are reported in many cancers as explained below.

Brain tumor

Recent studies on clinical samples showed increased levels of IGF1 as well as IGF-sequestering IGFBP3, IGFBP3-cleaving proteases (MMP and tPA) in CSF and IGF1R was identified as a potential target for clinical trials in high risk medulloblastoma [7]. Expression studies on glioma patient samples shown to have increased expression of IGF1R compares to normal brain tissue [8] and overexpression of IRS1 was reported in both malignant and benign brain tumors [9,10]. The overexpression of IRS1 as well as IRS2 in different cancers including medulloblastoma have reviewed earlier indicating its role in tumorigenesis [11,12]. Validation of genes related to the PI3k/Akt signaling pathway for genetic alteration and aberrant expression in a series of 103 glioblastomas showed overexpression of several genes including IRS2, AKT1 and subunits of Pi3k, but not IRS1 or AKT2 [13].

Head and neck cancer

Validation of IGF1R- α cytoplasmic/membranous overexpression showed it may serve as an independent adverse prognostic factor for recurrence and survival in patients with surgically resected squamous-cell carcinoma of the larynx [14] where a previous study showed IGFBP-3 expression, in a state of co-expression with IGF-1R, can predict poor prognosis in squamous cell carcinoma of the head and neck (SCCHN) patients [15]. Tissue level expression of IGF1R revealed an overexpression in both in primaries and regional lymph nodes of oropharyngeal and nasopharyngeal region [16] and in tongue cancer patients [17], which is capable of transmitting mitogenic signals to the neoplastic cells in response to IGF1. In HNSCC high IGF-1R expression and its association with HPV-negative status, advanced tumor stage and reduced overall and disease-specific survival [18] makes it a promising target for HPV negative HNSCC. In nasopharyngeal carcinoma, overexpression of IRS1 might be closely correlated with lymph node metastasis in NPC and positive expression of IRS-1 is postulated to be used as an independent biomarker for predicting lymph node metastasis of NPC [19] but there are contradicting results showing negative correlation of IRS-1 expression with tumor metastasis both in tissue samples and cell lines of HNSCC [20]. Over expression of IGF1R, IRS and MAPK1 was reports in a study where significant number of head and neck cancer patient samples showed the upregulation in tumor compared to the normal counterpart [21].

Breast cancer

Studies on benign and noninvasive breast lesions, IGF-1R expression was slightly increased in lesions that are hormonally driven (such as atypical ductal hyperplasia and columnar cells changes) and was significantly reduced in ER-negative lesions (such as apocrine metaplasia) which may explain partially lower response rate to neo adjuvant chemotherapy in luminal tumors [22]. IGF-1R expression was associated with

mammographic breast tissue density in patients aged >40 y and in premenopausal women, IGF-1R overexpression in breast cancer tissue was significantly associated with HER-2 positivity and poor disease free survival (DFS) [23].

In MCF-7 overexpression of IRS1 alone was capable of total loss of estrogen dependence for growth [24] and IRS1 showed a highly significant positive correlation and highly expressed in the less aggressive ductal carcinoma in situ (DCIS) tumors whereas elevated IRS2 expression was associated with high grade invasive ductal carcinomas (IDC) and positive correlation between ER α and IRS1 was hypothesized to respond favorably to chemotherapy [25] and in breast cancer progression [26]. Studies on transgenic mice, overexpression IRS1 and IRS2 in the mammary gland showed progressive mammary hyperplasia, tumorigenesis, and metastasis [27]. Role of IRS proteins in mammary gland development and in its function as well as in tumor formation and metastasis has reviewed earlier [28,29].

Lung cancer

Deregulation in IGF1R pathway and beneficial role of targeting that pathway is reviewed elsewhere [30]. Upregulation and expression of IGF-1 and IGF1R in NSCLC were correlated with tumor progression and patient prognosis [31]. Over expression of IGF1R protein in small cell lung cancer was reported to be correlated with the gene copy number in clinical samples [32] and an independent study on IGF-1R overexpression in squamous cell lung cancer displayed 79.2% of high IGF-1R staining [33]. Another study on NSCLC tissues revealed IGF1R protein expression and mRNA expression is higher in squamous cell carcinomas compared with other histologies which correlated with EGFR expression and high IGF1R gene copy number alone associates with better prognosis in surgically resected NSCLC [34].

Gastrointestinal cancers

Studies on wild type (WT) and pediatric GISTs (gastrointestinal stromal tumors) revealed that IGF1R is amplified and the protein is overexpressed which may be associated with oncogenesis [35] and the overexpression of IGF1R at protein and transcript was tightly SDHB (succinate dehydrogenase) deficiency [36]. Integrative genomic analysis showed enrichment of activation of IGF signaling, including overexpression of IGF2, downregulation of IGFBP3 and allelic losses of IGF2R and the study defined gene signature for IGF-1R activation finally leads to the Proliferation subclass of hepatocellular carcinoma [37]. Over expression of IGF2 as well as IGF1R in protein as well as in RNA level was involved in the pathogenesis of gastric cancer, probably by autocrine/paracrine stimulation of cell growth [38]. In patient samples with colorectal cancer, expression of IGF-1 and IGF-1R seems to increase with tumor size [39]. Expression studies of IGF1R in colorectal cancer patients revealed the fact that functional changes of the IGF-1R expression participate in the maintenance of progression of cancer rather than in its initial development [40].

Bone cancer

Studies on osteosarcoma showed higher mRNA expression and copy number in primary patient samples and xenograft samples compared with corresponding cell lines [41] and the

significant level of expression of IGF1R and the ligands were not correlated with metastatic lesions [42]. Studies on mouse mesenchymal stem cells (MSCs) and in osteosarcoma cells showed a precise regulation of IRS-1 expression level is critical for determining the differentiating capacity of these and that derangement of IRS-1 levels can be a critical step in OS transformation [43]. Genome-wide gene expression data of high-grade osteosarcoma biopsies and cell lines detected an overrepresentation of IGF1R signaling including downregulation of several IGF1R and further inhibition of IGF1R as well as IR/IGF1R signaling using small molecule inhibitor reduced the phosphorylation of IRS1 [44].

Prostate cancer

Over expression of IGF1R in primary prostate cancer compared with benign prostatic epithelium and the observed increase in IRS-1, a docking protein in IGF1R pathway, in a few sample, highlight the importance of the IGF1R pathway in prostate cancer biology [45]. IGF-1R analysis in PCa cells or clinical samples showed that T2E (MPRSS2-ERG (T2E) fusion protein) expression correlated with higher IGF-1R expression at mRNA and protein levels where ERG acts as a transcriptional activator for IGF1R [46]. Studies on PC cell line showed IGF-1 as a novel survival factor in prostate cancer, which induces a resistance to apoptosis mediated by the expression of the anti-apoptotic proteins Bcl⁻XL and Mcl⁻1 and is associated with increased phosphorylation of FAK [47]. An independent study on advanced, androgen-independent metastatic PC tumor samples, IGF1R and IRS-1 continued to express during the metastatic development of cancer [45] and in an *in vivo* study, upregulation of both IGF-1 and the IGF1R during progression to androgen-independent growth of tumor was observed [48].

Others

Wilms tumor which is a pediatric metanephric blastema, is observed to have overexpression of IGF1R related to the WT1, Wilms tumor gene product binding to IGF1R promoter [49]. In synovial sarcoma tumors, IGF1R expression check was done using Western blot and reverse transcriptase-PCR and the IGF1R detectable group was associated with a high incidence of lung metastases and high tumor cell proliferative rate, compared with the non-detectable group [50]. Molecular profiling studies on human adrenal tumors validated overexpression of 2 precarious components of the IGF signaling cascade (IGF2 and IGF-1R) and associated activation of the downstream effector, Akt. These results confirming critical role of IGF pathway in ACC pathogenesis [51]. In case of familial pheo/pgl, overexpressed IGF-1R was found to be associated with malignancy and such patients are hypothesized to have an increased risk of metastasis [52]. IGF1R upregulation in bladder cancer compared with non-malignant bladder especially increased transcript level of IGF1R in invasive (T2–4) tumors can relate its contribution to a propensity for invasion [53], but an independent experiment with patient as well as normal subjects showed no difference in free IGF1 level in urine samples [54]. Increased expression of IGF1R as well as IGF1 in chronic lymphocytic leukemia compared with

non CLL and the observation that use of TKI, inhibited the several signaling molecules via by blocking IGF1R-mediated signaling, makes this an ideal pathway for CLL treatment in future [55]. In a study on malignant adrenocortical tumor, the IGF1R overexpression was found to be independent of increased gene copy number, IGF1R polymorphisms and the abnormal expression of the microRNAs miR-100, miR-145, miR-375, and miR-126 [56]. Large cohort Genomic characterization of Malignant peripheral nerve sheath tumor (MPNST) identified a significant enrichment of copy number—an important events in the insulin-like growth factor 1 receptor (IGF1R) pathway, including frequent amplifications of the IGF1R gene itself [57]. IRS2 expression studies on a large variety of tumors [58] neurofibromas showed its elevated expression during its the progression to malignant peripheral nerve sheath tumors (MPNSTs) but it does not necessarily correlate with poor survival [59].

miR against IGF1R pathway

Small endogenous, non-coding microRNAs are validated to have significant role in diverse biologic processes which vary from development to different diseases including cancer. Deregulated miR expression was studied extensively in cancer elsewhere [60,61]. The ability of a single miR to regulate expression of multiple mRNAs makes further validation of it in therapy a promising area of research [62]. Independent studies have confirmed different miRs which can target single or multiple molecules in IGF1R pathway [21,63–118]. Existing validated data have shown miRs which can target multiple molecules in IGF1R pathway including IGF1R, which include miR-7, Let-7, miR-493, miR-155, miR-99, miR-145 etc. as shown in Fig. 1.

Perspective

IGF1 pathway has been extensively studied for the identification of deregulated miR which targets the members of this pathway [119]. Microarray as well as qRT-PCR studies have confirmed significant miRs in paired tumor samples as well as in cell lines in different cancer types. Regulation of miR transcription through IGF1R pathway and reciprocal regulation of IGF1R pathway by same miR is an indication of importance of controlled signaling pathways within cells. Reciprocal Regulation of miR and IGF1R pathway have been reported in different studies as miR-1 and Insulin-Like Growth Factor-1 Signal Transduction Cascade through Foxo3a [120], miR-99a via IGF1R/MAPK and IGF1R/AKT signaling [121,122], miR-493 expression through snail [21], miR-122 via C/EBP α [123]. These kinds of experimental evidences indicate the importance of existence miR mediated feedback loops in growth factor mediated signaling and their significance in normal functioning of cells. Use of miRs which can target multiple molecules in IGF1 signaling pathway can be used with successful delivery methods in different cancers where overexpression of multiple pathway molecules are evident and thereby reduce therapeutic resistance. Such therapy will reduce the recurrence of cancer and improve disease free survival of patient.

- [20] Luo X, Fan S, Huang W, et al. Downregulation of IRS-1 promotes metastasis of head and neck squamous cell carcinoma. *Oncol Rep.* 2012;28(2):659-67. doi:10.3892/or.2012.1846
- [21] Kumar AS, Jagadeeshan S, Pitani RS, et al. Snail modulated miR-493 forms a negative feedback loop with IGF1R pathway and blocks tumorigenesis. *Mol Cell Biol.* 2016;37(6):pii: e00510-16 MCB-00510. doi:10.1128/MCB.00510-16
- [22] Bhargava R, Beriwal S, McManus K, et al. Insulin-like growth factor receptor-1 (IGF-1R) expression in normal breast, proliferative breast lesions, and breast carcinoma. *Appl Immunohistochem Mol Morphol.* 2011;19(3):218-25. doi:10.1097/PAL.0b013e3181ffc58c
- [23] Sun WY, Yun HY, Song YJ, et al. Insulin-like growth factor 1 receptor expression in breast cancer tissue and mammographic density. *Mol Clin Oncol.* 2015;3(3):572-80. doi:10.3892/mco.2015.497
- [24] Surmacz E, Burgaud JL. Overexpression of insulin receptor substrate 1 (IRS-1) in the human breast cancer cell line MCF-7 induces loss of estrogen requirements for growth and transformation. *Clin Cancer Res.* 1995;1(11):1429-36
- [25] Porter HA, Perry A, Kingsley C, et al. IRS1 is highly expressed in localized breast tumors and regulates the sensitivity of breast cancer cells to chemotherapy, while IRS2 is highly expressed in invasive breast tumors. *Cancer Lett.* 2013;338(2):239-48. doi:10.1016/j.canlet.2013.03.030
- [26] Koda M, Sulowska M, Kanczuga-Koda L, et al. Expression of insulin receptor substrate 1 in primary breast cancer and lymph node metastases. *J Clin Pathol.* 2005;58(6):645-9. doi:10.1136/jcp.2004.022590
- [27] Dearth RK, Cui X, Kim HJ, et al. Mammary tumorigenesis and metastasis caused by overexpression of insulin receptor substrate 1 (IRS-1) or IRS-2. *Mol Cell Biol.* 2006;26(24):9302-14. doi:10.1128/MCB.00260-06
- [28] Dearth RK, Cui X, Kim HJ, et al. Oncogenic transformation by the signaling adaptor proteins insulin receptor substrate (IRS)-1 and IRS-2. *Cell Cycle.* 2007;6(6):705-13. doi:10.4161/cc.6.6.4035
- [29] Gibson SL, Ma Z, Shaw LM. Divergent roles for IRS-1 and IRS-2 in breast cancer metastasis. *Cell cycle.* 2007;6(6):631-7. doi:10.4161/cc.6.6.3987
- [30] Dziadziuszko R, Camidge DR, Hirsch FR. The insulin-like growth factor pathway in lung cancer. *J Thoracic Oncol.* 2008;3(8):815-8. doi:10.1097/JTO.0b013e31818180f5
- [31] Fu S, Tang H, Liao Y, et al. Expression and clinical significance of insulin-like growth factor 1 in lung cancer tissues and perioperative circulation from patients with non-small-cell lung cancer. *Curr Oncol.* 2016;23(1):12. doi:10.3747/co.23.2669
- [32] Badzio A, Wynes MW, Dziadziuszko R, et al. Increased insulin-like growth factor 1 receptor protein expression and gene copy number in small cell lung cancer. *J Thoracic Oncol.* 2010;5(12):1905-11. doi:10.1097/JTO.0b013e3181f38f57
- [33] Gong Y, Yao E, Shen R, et al. High expression levels of total IGF-1R and sensitivity of NSCLC cells in vitro to an anti-IGF-1R antibody (R1507). *PLoS One.* 2009;4(10):e7273. doi:10.1371/journal.pone.0007273
- [34] Dziadziuszko R, Merrick DT, Witt SE, et al. Insulin-like growth factor receptor 1 (IGF1R) gene copy number is associated with survival in operable non-small-cell lung cancer: a comparison between IGF1R fluorescent in situ hybridization, protein expression, and mRNA expression. *J Clin Oncol.* 2010;28(13):2174-80. doi:10.1200/JCO.2009.24.6611
- [35] Tarn C, Rink L, Merkel E, et al. Insulin-like growth factor 1 receptor is a potential therapeutic target for gastrointestinal stromal tumors. *Proc Natl Acad Sci.* 2008;105(24):8387-92. doi:10.1073/pnas.0803383105
- [36] Belinsky MG, Rink L, Flieder DB, et al. Over-expression of IGF1R and frequent mutational inactivation of SDHA in Wild-type SDHB-negative gastrointestinal stromal tumors. *Genes Chromosomes Cancer.* 2013;52(2):214. doi:10.1002/gcc.22023
- [37] Tovar V, Alsinet C, Villanueva A, et al. IGF activation in a molecular subclass of hepatocellular carcinoma and pre-clinical efficacy of IGF-1R blockade. *J Hepatol.* 2010;52(4):550-9. doi:10.1016/j.jhep.2010.01.015
- [38] Pavelić K, Kolak T, Kapitanović S, et al. Gastric cancer: the role of insulin-like growth factor 2 (IGF 2) and its receptors (IGF 1R and IGF 2R). *J Pathol.* 2003;201(3):430-8. doi:10.1002/path.1465
- [39] Shiratsuchi I, Akagi Y, Kawahara A, et al. Expression of IGF-1 and IGF-1R and their relation to clinicopathological factors in colorectal cancer. *Anticancer Res.* 2011;31(7):2541-5
- [40] Karakolev I, Stanilov N, Miteva L, et al. Expression of insulin-like growth factor-1 receptor mRNA in colorectal carcinoma patients. *Biotechnol Biotechnological Equipment.* 2012;26(sup1):89-95. doi:10.5504/50YRTIMB.2011.0017
- [41] Cao Y, Roth M, Piperdi S, et al. Insulin-like growth factor 1 receptor and response to anti-IGF1R antibody therapy in osteosarcoma. *PLoS One.* 2014;9(8):e106249. doi:10.1371/journal.pone.0106249
- [42] Bell RS. Expression of insulin-like growth factor receptor, IGF-1, and IGF-2 in primary and metastatic osteosarcoma. *J Sur Oncol.* 1998;69:21-7. doi:10.1002/(SICI)1096-9098(199809)69:1%3c21::AID-JSO5%3e3.0.CO;2-M
- [43] Contaldo C, Myers TJ, Zucchini C, et al. Expression levels of insulin receptor substrate-1 modulate the osteoblastic differentiation of mesenchymal stem cells and osteosarcoma cells. *Growth Factors.* 2014;32(1):41-52. doi:10.3109/08977194.2013.870168
- [44] Kuijjer ML, Peterse EF, van den Akker BE, et al. IR/IGF1R signaling as potential target for treatment of high-grade osteosarcoma. *BMC Cancer.* 2013;13(1):245. doi:10.1186/1471-2407-13-245
- [45] Hellawell GO, Turner GD, Davies DR, et al. Expression of the type 1 insulin-like growth factor receptor is up-regulated in primary prostate cancer and commonly persists in metastatic disease. *Cancer Res.* 2002;62(10):2942-50
- [46] Mancarella C, Casanova-Salas I, Calatrava A, et al. ERG deregulation induces IGF-1R expression in prostate cancer cells and affects sensitivity to anti-IGF-1R agents. *Oncotarget.* 2015;6(18):16611-22. doi:10.18632/oncotarget.3425
- [47] Watson RW, O'Brien F, Coffey RN, et al. Insulin-like growth factor-1 alters apoptotic signalling in prostate cancer. *Prostate Cancer Prostatic Dis.* 2000;3(4):S42. doi:10.1038/sj.pcan.4500468
- [48] Nickerson T, Chang F, Lorimer D, et al. In vivo progression of LAPC-9 and LNCaP prostate cancer models to androgen independence is associated with increased expression of insulin-like growth factor I (IGF-I) and IGF-I receptor (IGF-IR). *Cancer Res.* 2001;61(16):6276-80
- [49] Werner H, Re GG, Drummond IA, et al. Increased expression of the insulin-like growth factor I receptor gene, IGF1R, in Wilms tumor is correlated with modulation of IGF1R promoter activity by the WT1 Wilms tumor gene product. *Proc Natl Acad Sci.* 1993;90(12):5828-32. doi:10.1073/pnas.90.12.5828
- [50] Xie Y, Skytting B, Nilsson G, et al. Expression of insulin-like growth factor-1 receptor in synovial sarcoma. *Cancer Res.* 1999;59(15):3588-91
- [51] Barlaskar FM, Spalding AC, Heaton JH, et al. Preclinical targeting of the type I insulin-like growth factor receptor in adrenocortical carcinoma. *J Clin Endocrinol Metab.* 2009;94(1):204-12. doi:10.1210/jc.2008-1456
- [52] Fernandez MC, Martin A, Venara M, et al. Overexpression of the insulin-like growth factor 1 receptor (IGF-1R) is associated with malignancy in familial pheochromocytomas and paragangliomas. *Clin Endocrinol.* 2013;79(5):623-30. doi:10.1111/cen.12205
- [53] Rochester MA, Patel N, Turney BW, et al. The type 1 insulin-like growth factor receptor is over-expressed in bladder cancer. *BJU Int.* 2007;100(6):1396-401. doi:10.1111/j.1464-410X.2007.06931.x
- [54] Serel TA, Turan T, Soyupek S, et al. Urine and serum free IGF-1 levels in patients with bladder cancer: a brief report. *Urol Res.* 2003;31(5):297-9. doi:10.1007/s00240-003-0335-0
- [55] Yaktapour N, Übelhart R, Schüller J, et al. Insulin-like growth factor-1 receptor (IGF1R) as a novel target in chronic lymphocytic leukemia. *Blood.* 2013;122(9):1621-33. doi:10.1182/blood-2013-02-484386
- [56] Ribeiro TC, Jorge AA, Almeida MQ, et al. Amplification of the insulin-like growth factor 1 receptor gene is a rare event in

- adrenocortical adenocarcinomas: searching for potential mechanisms of overexpression. *Biomed Res Int.* 2014;2014:936031. doi:10.1155/2014/936031
- [57] Yang J, Ylipää A, Sun Y, et al. Genomic and molecular characterization of malignant peripheral nerve sheath tumor identifies the IGF1R pathway as a primary target for treatment. *Clin Cancer Res.* 2011;17(24):7563-73. doi:10.1158/1078-0432.CCR-11-1707
- [58] Chang Q, Li Y, White MF, et al. Constitutive activation of insulin receptor substrate 1 is a frequent event in human tumors. *Cancer Res.* 2002;62(21):6035-8
- [59] Shaw CM, Grobmyer SR, Ucar DA, et al. Elevated expression of IRS2 in the progression from neurofibroma to malignant peripheral nerve sheath tumor. *Anticancer Res.* 2012;32(2):439-43
- [60] Rupaimoole R, Calin GA, Lopez-Berestein G, et al. miRNA deregulation in cancer cells and the tumor microenvironment. *Cancer Discover.* 2016;6(3):235-46. doi:10.1158/2159-8290.CD-15-0893
- [61] Iorio MV, Croce CM. Causes and consequences of microRNA dysregulation. *Cancer J (Sudbury, Mass.).* 2012;18(3):215. doi:10.1097/PPO.0b013e318250c001
- [62] Berindan-Neagoe I, Monroig PD, Pasculli B, et al. MicroRNAome genome: a treasure for cancer diagnosis and therapy. *CA Cancer J Clin.* 2014;64(5):311-36. doi:10.3322/caac.21244
- [63] Wu X, Xia M, Chen D, et al. Profiling of downregulated blood-circulating miR-150-5p as a novel tumor marker for cholangiocarcinoma. *Tumor Biol.* 2016;37(11):15019-29. doi:10.1007/s13277-016-5313-6
- [64] Law PT, Ching AK, Chan AW, et al. MiR-145 modulates multiple components of the insulin-like growth factor pathway in hepatocellular carcinoma. *Carcinogenesis.* 2012;33(6):1134-41. doi:10.1093/carcin/bgs130
- [65] Tardif G, Hum D, Pelletier JP, et al. Regulation of the IGF1R and MMP-13 genes by the microRNAs miR-140 and miR-27a in human osteoarthritic chondrocytes. *BMC Musculoskelet Disord.* 2009;10(1):148. doi:10.1186/1471-2474-10-148
- [66] El Tayebi HM, Waly AA, Assal RA, et al. Transcriptional activation of the IGF-II/IGF-1R axis and inhibition of IGF1R-3 by miR-155 in hepatocellular carcinoma. *Oncol Lett.* 2015;10(5):3206-12. doi:10.3892/ol.2015.3725
- [67] Xu Y, Ouyang X, Chen X, et al. Inhibition effect of miR-7 on proliferation and metastasis of Hep-2 laryngeal carcinoma cells. *Chinese Clinical Oncology.* 2015;3.
- [68] Okamoto K, Ishiguro T, Midorikawa Y, et al. miR-493 induction during carcinogenesis blocks metastatic settlement of colon cancer cells in liver. *EMBO J.* 2012;31(7):1752-63. doi:10.1038/emboj.2012.25
- [69] Ying W, Tseng A, Chang RC, et al. miR-150 regulates obesity-associated insulin resistance by controlling B cell functions. *Sci Rep.* 2016;6:20176. doi:10.1038/srep20176
- [70] Xu F, Pang L, Cai X, et al. let-7-repressed Shc translation delays replicative senescence. *Aging cell.* 2014;13(1):185-92. doi:10.1111/accel.12176
- [71] Park SY, Lee JH, Ha M, et al. miR-29 miRNAs activate p53 by targeting p85 α and CDC42. *Nat Struct Mol Biol.* 2009;16(1):23-9. doi:10.1038/nsmb.1533
- [72] Yang Y, Liu L, Zhang Y, et al. MiR-503 targets PI3K p85 and IKK- β and suppresses progression of non-small cell lung cancer. *Int J Cancer.* 2014;135(7):1531-42. doi:10.1002/ijc.28799
- [73] Tsukamoto Y, Nakada C, Noguchi T, et al. MicroRNA-375 is downregulated in gastric carcinomas and regulates cell survival by targeting PDK1 and 14-3-3 ζ . *Cancer Res.* 2010;70(6):2339-49. doi:10.1158/0008-5472.CAN-09-2777
- [74] Liu Z, Jiang Z, Huang J, et al. miR-7 inhibits glioblastoma growth by simultaneously interfering with the PI3K/ATK and Raf/MEK/ERK pathways. *Int J Oncol.* 2014;44(5):1571-80. doi:10.3892/ijo.2014.2322
- [75] Fang B, Zhu J, Wang Y, et al. MiR-454 inhibited cell proliferation of human glioblastoma cells by suppressing PDK1 expression. *Biomed Pharmacother.* 2015;75:148-52. doi:10.1016/j.biopha.2015.07.029
- [76] Liu Y, Yang K, Sun X, et al. MiR-138 suppresses airway smooth muscle cell proliferation through the PI3K/AKT signaling pathway by targeting PDK1. *Exp Lung Res.* 2015;41(7):363-9. doi:10.3109/01902148.2015.1041581
- [77] Wang Y, Zhang Z, Wang H, et al. miR-138-1* regulates aflatoxin B1-induced malignant transformation of BEAS-2B cells by targeting PDK1. *Arch Toxicol.* 2016;90(5):1239-49. doi:10.1007/s00204-015-1551-4
- [78] Huang X, Shen Y, Liu M, et al. Quantitative proteomics reveals that miR-155 regulates the PI3K-AKT pathway in diffuse large B-cell lymphoma. *Am J Pathol.* 2012;181(1):26-33. doi:10.1016/j.ajpath.2012.03.013
- [79] Zhao HJ, Ren LL, Wang ZH, et al. MiR-194 deregulation contributes to colorectal carcinogenesis via targeting AKT2 pathway. *Theranostics.* 2014;4(12):1193-208. doi:10.7150/thno.8712
- [80] Ichimura A, Ruike Y, Terasawa K, et al. MicroRNA-34a inhibits cell proliferation by repressing mitogen-activated protein kinase kinase 1 during megakaryocytic differentiation of K562 cells. *Mol Pharmacol.* 2010;77(6):1016-24. doi:10.1124/mol.109.063321
- [81] Pang L, You L, Ji C, et al. MiR-1275 inhibits adipogenesis via ELK1 and its expression decreases in obese subjects. *J Mol Endocrinol.* 2016;57(1):33-43. doi:10.1530/JME-16-0007
- [82] Bianchi N, Finotti A, Ferracin M, et al. Increase of microRNA-210, decrease of raptor gene expression and alteration of mammalian target of rapamycin regulated proteins following mithramycin treatment of human erythroid cells. *PloS One.* 2015;10(4):e0121567. doi:10.1371/journal.pone.0121567
- [83] Guo C, Sah JF, Beard L, et al. The noncoding RNA, miR-126, suppresses the growth of neoplastic cells by targeting phosphatidylinositol 3-kinase signaling and is frequently lost in colon cancers. *Genes Chromosomes Cancer.* 2008;47(11):939-46. doi:10.1002/gcc.20596
- [84] He W, Feng L, Xia D, et al. MiR-374a promotes the proliferation of human osteosarcoma by downregulating FOXO1 expression. *Int J Clin Exp Med.* 2015;8(3):3482; PMID:PMC4443073
- [85] Xu Y, Zhao S, Cui M, et al. Down-regulation of microRNA-135b inhibited growth of cervical cancer cells by targeting FOXO1. *Int J Clin Exp Pathol.* 2015;8(9):10294; PMID:PMC4637552
- [86] Mao XP, Zhang LS, Huang B, et al. Mir-135a enhances cellular proliferation through post-transcriptionally regulating PHLPP2 and FOXO1 in human bladder cancer. *J Transl Med.* 2015;13(1):86. doi:10.1186/s12967-015-0438-8
- [87] Lian R, Lu B, Jiao L, et al. MiR-132 plays an oncogenic role in laryngeal squamous cell carcinoma by targeting FOXO1 and activating the PI3K/AKT pathway. *Euro J Pharmacol.* 2016;792:1-6. doi:10.1016/j.ejphar.2016.10.015
- [88] Cimmino A, Calin GA, Fabbri M, et al. miR-15 and miR-16 induce apoptosis by targeting BCL2. *Proc Natl Acad Sci U S A.* 2005;102(39):13944-9. doi:10.1073/pnas.0506654102
- [89] Lin X, Guan H, Huang Z, et al. Downregulation of Bcl-2 expression by miR-34a mediates palmitate-induced Min6 cells apoptosis. *J Diabetes Res.* 2014;2014:258695. doi:10.1155/2014/258695
- [90] Zhu P, Zhang J, Zhu J, et al. MiR-429 induces gastric carcinoma cell apoptosis through Bcl-2. *Cell Physiol Biochem.* 2015;37(4):1572-80. doi:10.1159/000438524
- [91] Zhang Y, Huang F, Wang J, et al. MiR-15b mediates liver cancer cells proliferation through targeting BCL-2. *Int J Clin Exp Pathol.* 2015;8(12):15677; PMID:PMC4730050
- [92] Han R, Ji X, Rong R, et al. MiR-449a regulates autophagy to inhibit silica-induced pulmonary fibrosis through targeting Bcl2. *J Mol Med.* 2016;94(11):1267-79. doi:10.1007/s00109-016-1441-0
- [93] Lin YC, Lin JF, Tsai TF, et al. Tumor suppressor miRNA-204-5p promotes apoptosis by targeting BCL2 in prostate cancer cells. *Asian J Surg.* 2016; pii:S1015-9584(16)30173-7. doi:10.1016/j.asjsur.2016.07.001
- [94] Li Q, Liang X, Wang Y, et al. miR-139-5p inhibits the epithelial-mesenchymal transition and enhances the chemotherapeutic sensitivity of colorectal cancer cells by downregulating BCL2. *Sci Rep.* 2016;6:27157. doi:10.1038/srep27157
- [95] Liu DZ, Zhang HY, Long XL, et al. MiR-150 promotes prostate cancer stem cell development via suppressing p27Kip1. *Eur Rev Med Pharmacol Sci.* 2015;19(22):4344-52

- [96] Lu K, Wang J, Song Y, et al. miRNA-24-3p promotes cell proliferation and inhibits apoptosis in human breast cancer by targeting p27Kip1. *Oncol Rep.* 2015;34(2):995-1002. doi:10.3892/or.2015.4025
- [97] Wang X, Shen E, Wang Y, et al. MiR-196a regulates high glucose-induced mesangial cell hypertrophy by targeting p27kip1. *J Lab Autom.* 2015;20(4):491-9. doi:10.1177/2211068215569055
- [98] Wang DD, Li J, Sha HH, et al. miR-222 confers the resistance of breast cancer cells to Adriamycin through suppression of p27 kip1 expression. *Gene.* 2016;590(1):44-50. doi:10.1016/j.gene.2016.06.013
- [99] Li Z, Xu Z, Xie Q, et al. miR-1303 promotes the proliferation of neuroblastoma cell SH-SY5Y by targeting GSK3 β and SFRP1. *Biomed Pharmacother.* 2016;83:508-13. doi:10.1016/j.biopha.2016.07.010
- [100] Guo LM, Pu Y, Han Z, et al. MicroRNA-9 inhibits ovarian cancer cell growth through regulation of NF- κ B1. *FEBS J.* 2009;276(19):5537-46. doi:10.1111/j.1742-4658.2009.07237.x
- [101] Zhao XD, Lu YY, Guo H, et al. MicroRNA-7/NF- κ B signaling regulatory feedback circuit regulates gastric carcinogenesis. *J Cell Biol.* 2015;210(4):613-27. doi:10.1083/jcb.201501073
- [102] Gu R, Liu N, Luo S, et al. MicroRNA-9 regulates the development of knee osteoarthritis through the NF-kappaB1 pathway in chondrocytes. *Medicine.* 2016;95(36):e4315. doi:10.1097/MD.00000000000004315
- [103] Zhang J, Zhang H, Liu J, et al. miR-30 inhibits TGF- β 1-induced epithelial-to-mesenchymal transition in hepatocyte by targeting Snail1. *Biochem Biophys Res Commun.* 2012;417(3):1100-5. doi:10.1016/j.bbrc.2011.12.121
- [104] Siemens H, Jackstadt R, Hüntgen S, et al. miR-34 and SNAIL form a double-negative feedback loop to regulate epithelial-mesenchymal transitions. *Cell Cycle.* 2011;10(24):4256-71. doi:10.4161/cc.10.24.18552
- [105] Zhou Q, Yang M, Lan H, et al. miR-30a Negatively Regulates TGF- β 1-Induced Epithelial-Mesenchymal Transition and Peritoneal Fibrosis by Targeting Snail. *Am J Pathol.* 2013;183(3):808-19. doi:10.1016/j.ajpath.2013.05.019
- [106] Suzuki T, Mizutani K, Minami A, et al. Suppression of the TGF- β 1-induced protein expression of SNAIL and N-cadherin by miR-199a. *Genes Cells.* 2014;19(9):667-75. doi:10.1111/gtc.12166
- [107] Muraoka N, Yamakawa H, Miyamoto K, et al. MiR-133 promotes cardiac reprogramming by directly repressing Snail and silencing fibroblast signatures. *EMBO J.* 2014;33(14):1565-81. doi:10.15252/embj.201387605
- [108] Qin H, Sha J, Jiang C, et al. miR-122 inhibits metastasis and epithelial-mesenchymal transition of non-small-cell lung cancer cells. *OncoTargets Ther.* 2015;8:3175. doi:10.2147/OTT.S91696
- [109] Yuan CT, Li XX, Cheng QJ, et al. MiR-30a regulates the atrial fibrillation-induced myocardial fibrosis by targeting snail 1. *Int J Clin Exp Pathol.* 2015;8(12):15527; PMID:PMC4730035
- [110] Bai Z, Sun J, Wang X, et al. MicroRNA-153 is a prognostic marker and inhibits cell migration and invasion by targeting SNAIL in human pancreatic ductal adenocarcinoma. *Oncol Rep.* 2015;34(2):595-602. doi:10.3892/or.2015.4051
- [111] Wang Z, Liu C. MiR-153 regulates metastases of gastric cancer through Snail. *Tumor Biol.* 2015;2:1-7. doi:10.1007/s13277-015-3846-8
- [112] Ye Z, Zhao L, Li J, et al. miR-30d Blocked Transforming Growth Factor β 1-Induced Epithelial-Mesenchymal Transition by Targeting Snail in Ovarian Cancer Cells. *Int J Gynecol Cancer.* 2015;25(9):1574-81. doi:10.1097/IGC.0000000000000546
- [113] Zuo QF, Cao LY, Yu T, et al. MicroRNA-22 inhibits tumor growth and metastasis in gastric cancer by directly targeting MMP14 and Snail. *Cell Death Dis.* 2015;6(11):e2000. doi:10.1038/cddis.2015.297
- [114] Zhang YF, Yu Y, Song WZ, et al. miR-410-3p suppresses breast cancer progression by targeting Snail. *Oncol Rep.* 2016;36(1):480-6. doi:10.3892/or.2016.4828
- [115] Dong P, Xiong Y, Watari H, et al. MiR-137 and miR-34a directly target Snail and inhibit EMT, invasion and sphere-forming ability of ovarian cancer cells. *J Exp Clin Cancer Res.* 2016;35(1):132. doi:10.1186/s13046-016-0415-y
- [116] Zuo J, Wang D, Shen H, et al. MicroRNA-153 inhibits tumor progression in esophageal squamous cell carcinoma by targeting SNAIL. *Tumor Biol.* 2016;13:1-6. doi:10.1007/s13277-016-5427-x
- [117] Zhang X, Zhang T, Yang K, et al. miR-486-5p suppresses prostate cancer metastasis by targeting snail and regulating epithelial-mesenchymal transition. *OncoTargets Ther.* 2016;9:6909. doi:10.2147/OTT.S117338
- [118] Wang K, Jin W, Jin P, et al. miR-211-5p suppresses metastatic behavior by targeting SNAIL in Renal Cancer. *Mol Cancer Res.* 2017;15(4):448-56. doi:10.1158/1541-7786.MCR-16-0288
- [119] Jung HJ, Suh Y. Regulation of IGF-1 signaling by microRNAs. *Front Genet.* 2015;5:472. doi:10.3389/fgene.2014.00472
- [120] Elia L, Contu R, Quintavalle M, et al. Reciprocal regulation of microRNA-1 and insulin-like growth factor-1 signal transduction cascade in cardiac and skeletal muscle in physiological and pathological conditions. *Circulation.* 2009;120(23):2377-85. doi:10.1161/CIRCULATIONAHA.109.879429
- [121] Yen YC, Shiah SG, Chu HC, Hsu YM, et al. Reciprocal regulation of microRNA-99a and insulin-like growth factor I receptor signaling in oral squamous cell carcinoma cells. *Mol Cancer.* 2014;13(1):6. doi:10.1186/1476-4598-13-6
- [122] Lerman G, Avivi C, Mardoukh C, et al. MiRNA expression in psoriatic skin: reciprocal regulation of hsa-miR-99a and IGF-1R. *PLoS One.* 2011;6(6):e20916. doi:10.1371/journal.pone.0020916
- [123] Zeng C, Wang R, Li D, et al. A novel GSK-3 β -C/EBP α -miR-122-insulin-like growth factor 1 receptor regulatory circuitry in human hepatocellular carcinoma. *Hepatology.* 2010;52(5):1702-12. doi:10.1002/hep.23875